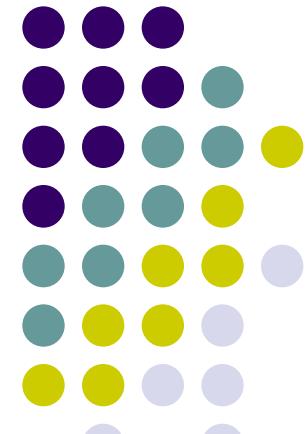
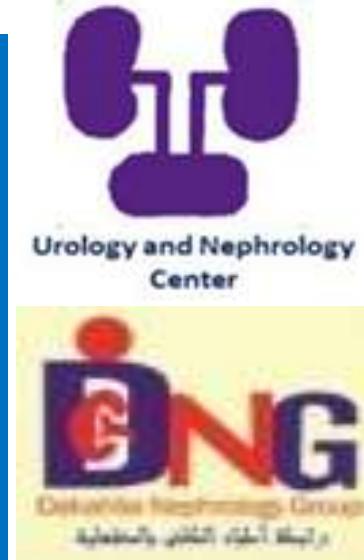


# Autophagy and The Aging Immune System

**Hussein Sheashaa, MD, FACP**

Professor of Nephrology, Urology and Nephrology Center and Director of  
Medical E-Learning Unit, Mansoura University and Executive Director of ESNT-  
Virtual Academy: <http://lms.mans.edu.eg/esnt/>



7- 8 October 2015  
**Egyptian Transplantation Society**  
**2nd Organ Transplantation Congress**  
Optimizing The Clinical Practice  
Cairo Marriott

# Focus of The Talk

- **Introduction**
- **Types and crosstalks**
- **Dysfunction and consequences**
- **Autophagy, immunity and immunosenescence**
- **Autophagy and mTOR**
- **Autophagy and transplantation**
- **Regulation**

Autophagy pres 7.10.2015

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multiple noxious stimuli in  
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Wen-jian Lin, et al.  
Volume 10, Issue 10, 2014



### The Nobel Prize in Physiology or Medicine 1974

Albert Claude, Christian de Duve, George E. Palade

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## Christian de Duve - Facts



Christian de Duve

**Born:** 2 October 1917, Thames Ditton, United Kingdom

**Died:** 4 May 2013, Nethen, Belgium

**Affiliation at the time of the award:** Rockefeller University, New York, NY, USA, Université Catholique de Louvain, Louvain, Belgium

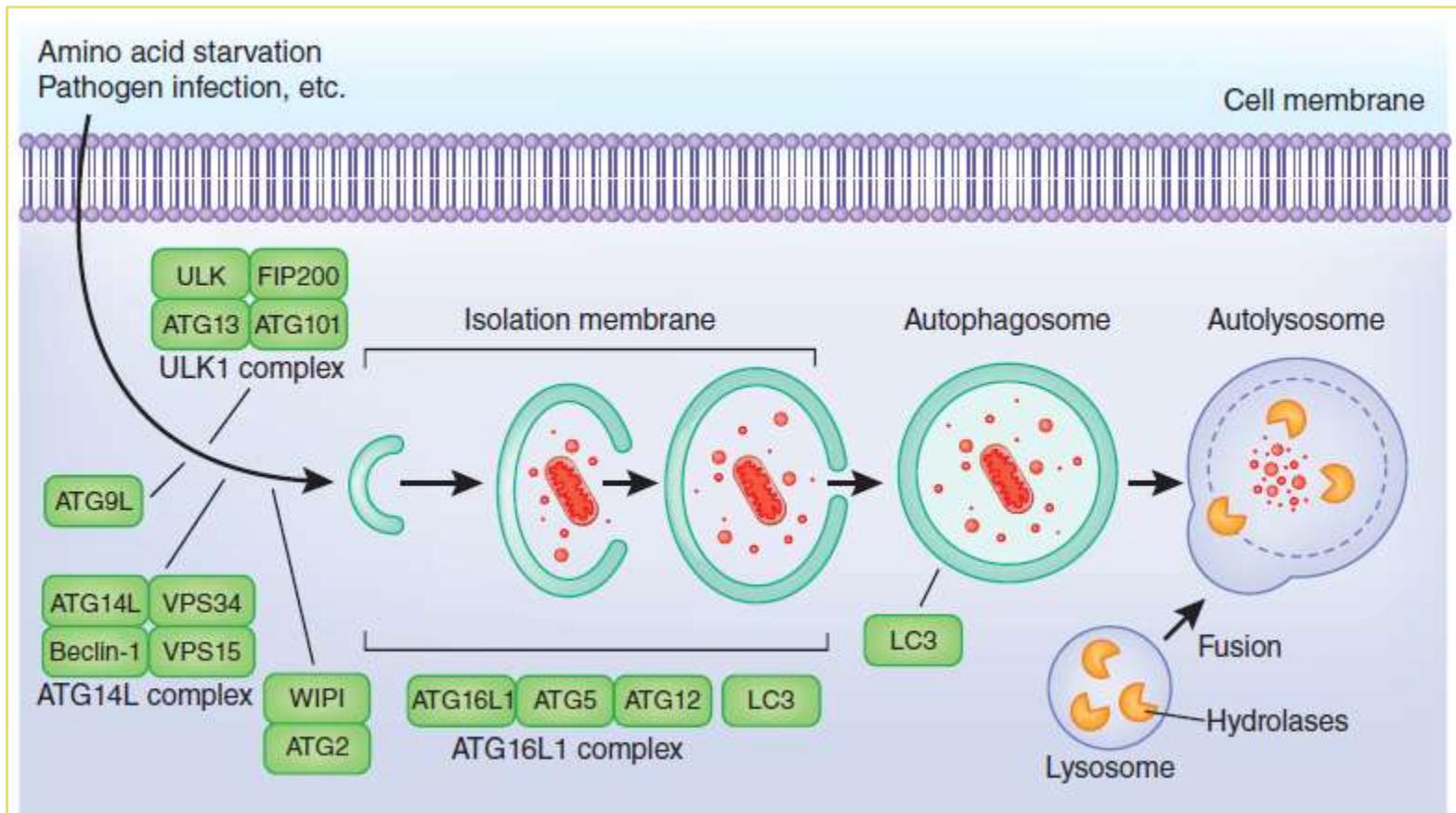
**Prize motivation:** "for their discoveries concerning the structural and functional organization of the cell"

**Field:** cell physiology

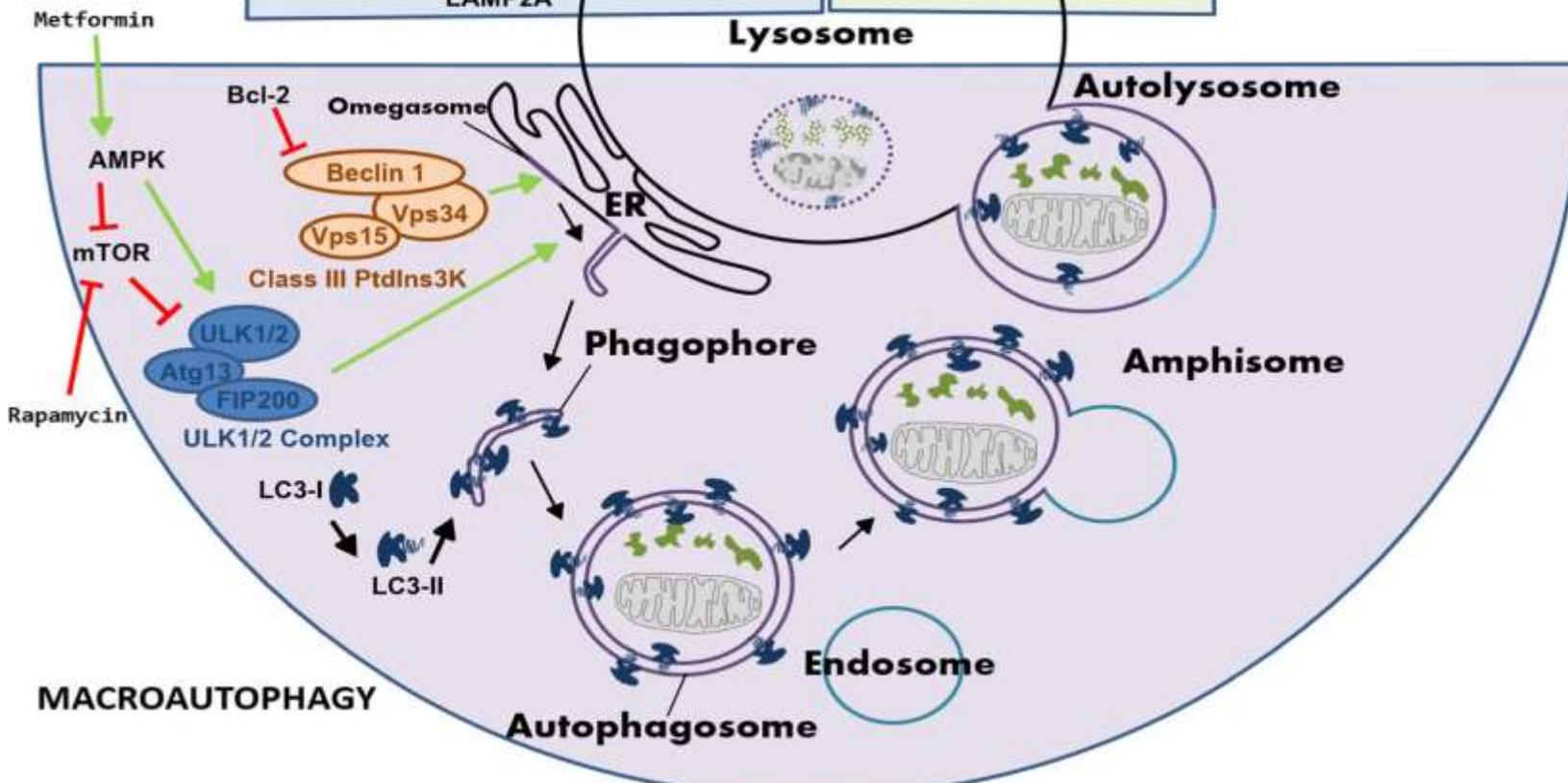
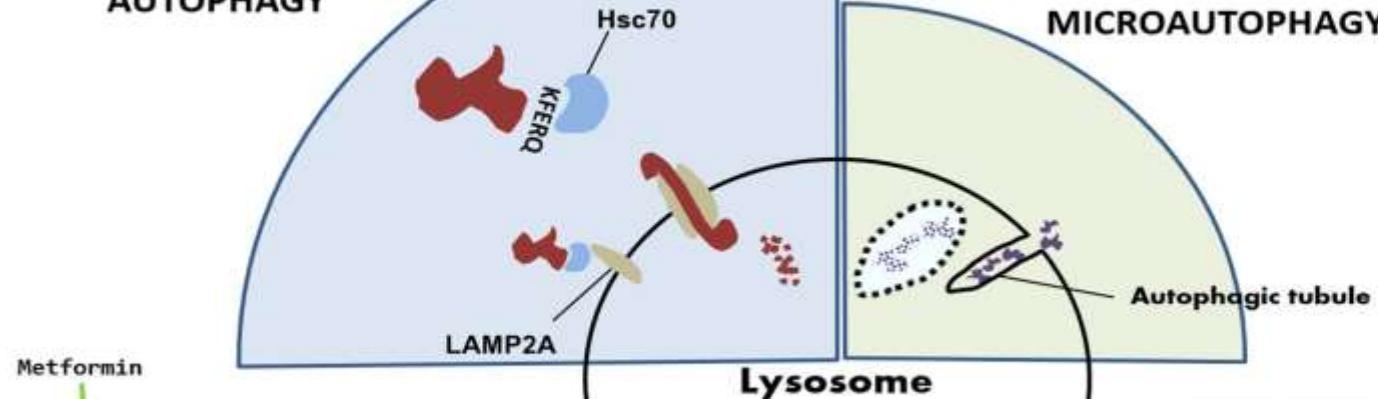
(2 October 1917 – 4 May 2013)

# Autophagy Types and Crosstalks

# Macroautophagy

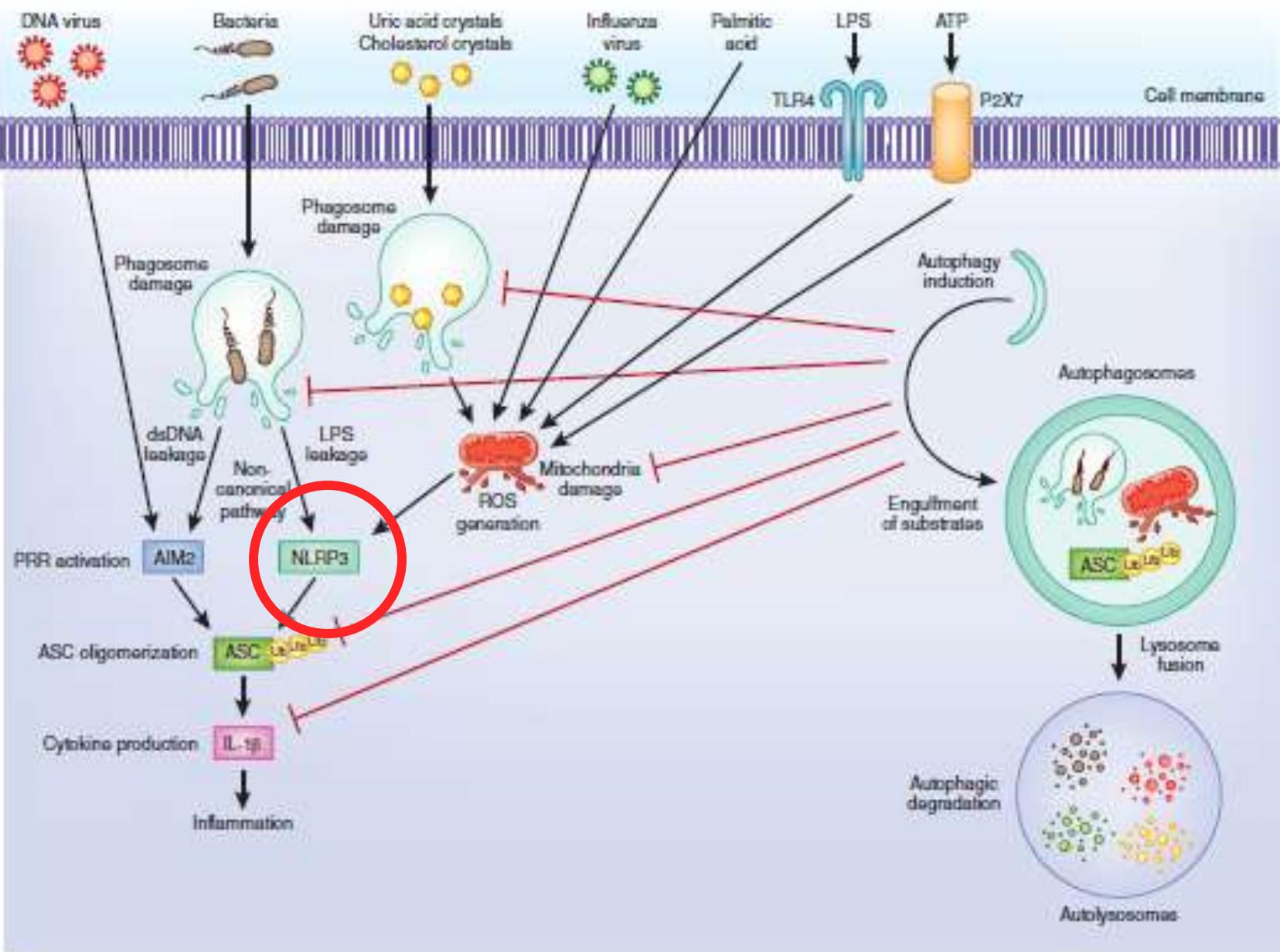


## CHAPERONE-MEDIATED AUTOPHAGY



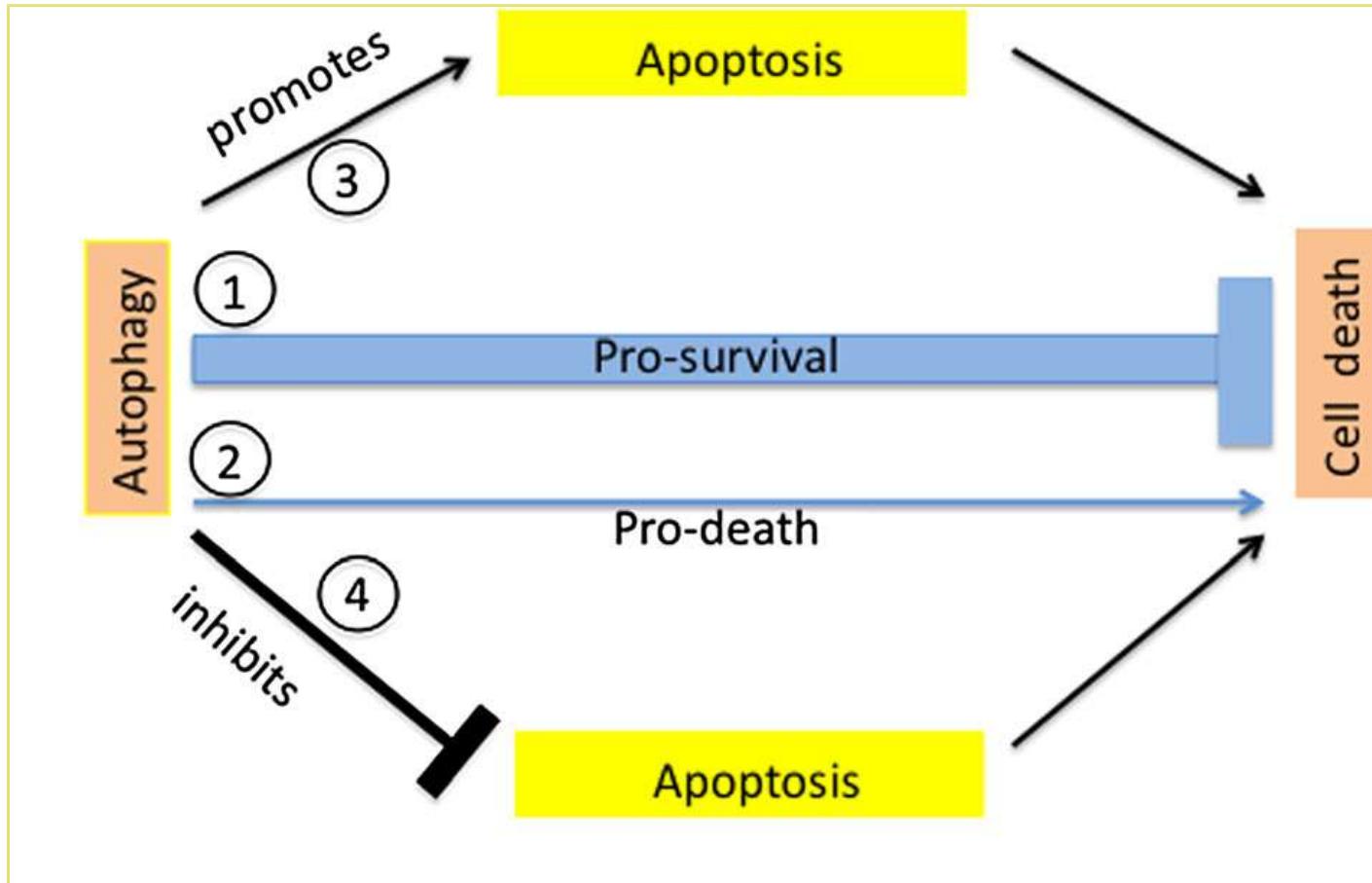


Urology and Nephrology Center



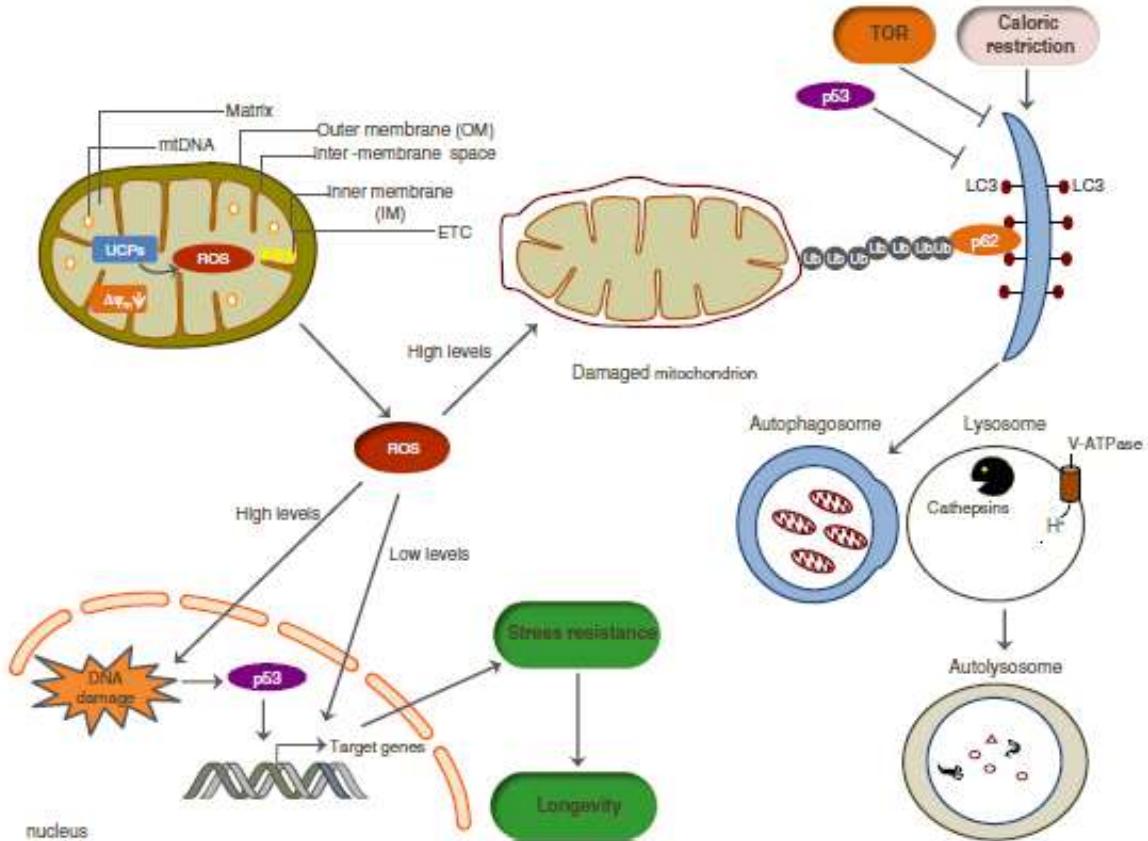
VOLUME 16 NUMBER 10 OCTOBER 2015 NATURE IMMUNOLOGY

# Autophagy and Apoptosis: Crosstalk



# Autophagy and Apoptosis: What Is The Difference?

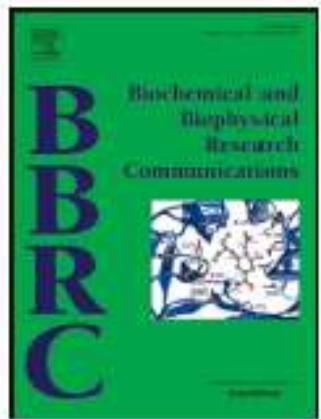
	Apoptosis	(Macro)autophagy	Autophagy- dependent cell death
Degradation of	Whole cell	Cellular components	Whole cell
Morphological features	Cell shrinkage Chromatin condensation Plasma membrane blebbing, preserved integrity Nuclear compaction and fragmentation Formation of apoptotic bodies		Minor changes to chromatin Plasma membrane rupture Minor changes to nucleus
		Autophagosomes Autolysosomes	Autophagosomes Autolysosomes
Interacting molecules	Bcl-2  Bcl-2 + BH3 mimetics  Caspases  Bcl-2  Bcl-XL	Beclin 1 / Class III PtdIns3K interaction  Beclin 1 / Class III PtdIns3K interaction  Atg proteins Beclin 1  Unconjugated Atg12  Atg12 – Atg3 complex	
Key molecules / regulators	TNF superfamily members Caspase proteases Bcl-2 proteins	Atg proteins Beclin 1 / Class III PtdIns3K ULK1/2 mTOR LC3	



# Accepted Manuscript

Autophagic response to cell culture stress in pluripotent stem cells

Sian Gregory, Sushma Swamy, Zoe Hewitt, Andrew Wood, Richard Weightman,  
Harry Moore



PII: S0006-291X(15)30597-0

DOI: [10.1016/j.bbrc.2015.09.080](https://doi.org/10.1016/j.bbrc.2015.09.080)

Reference: YBBRC 34580

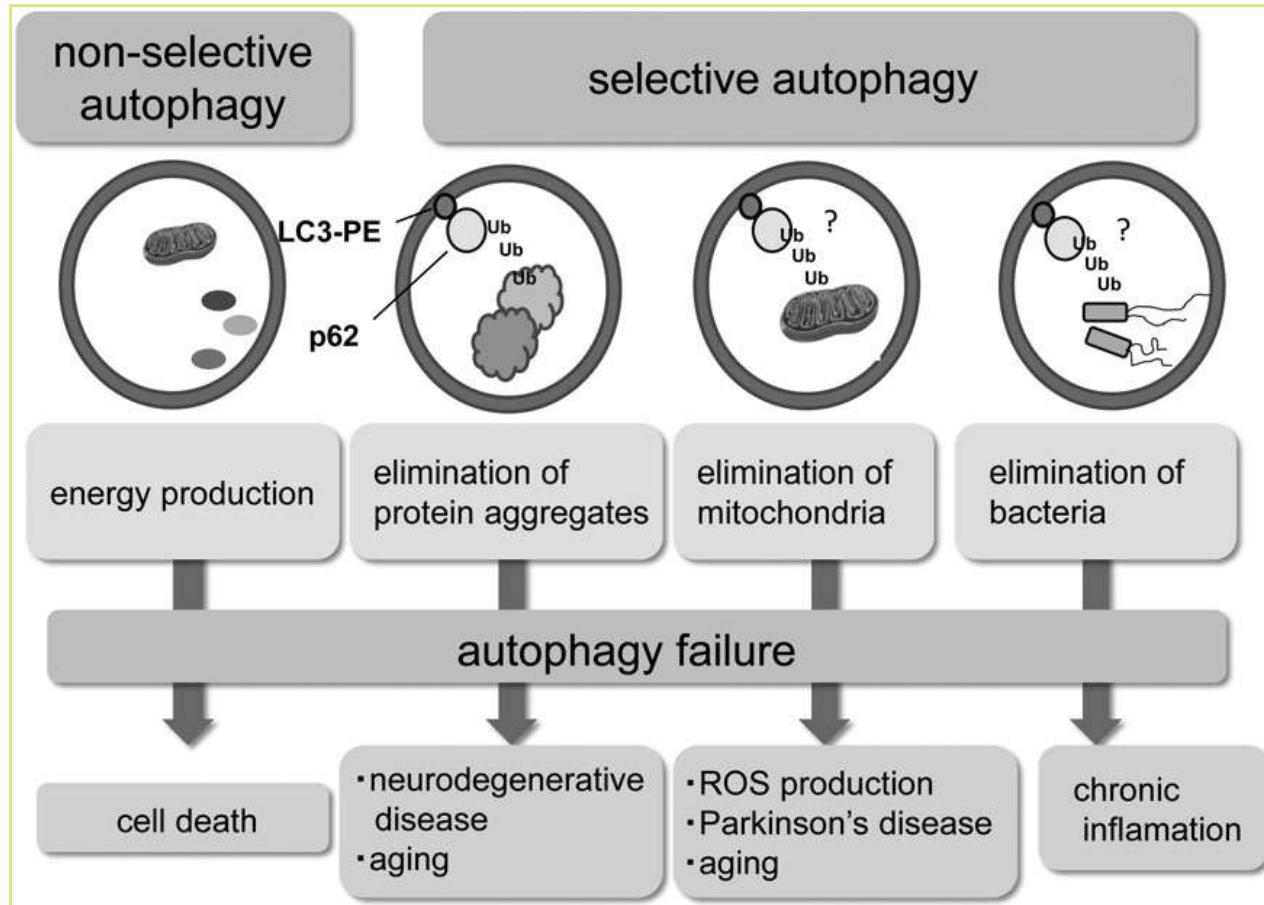
To appear in: *Biochemical and Biophysical Research Communications*

Received Date: 12 August 2015

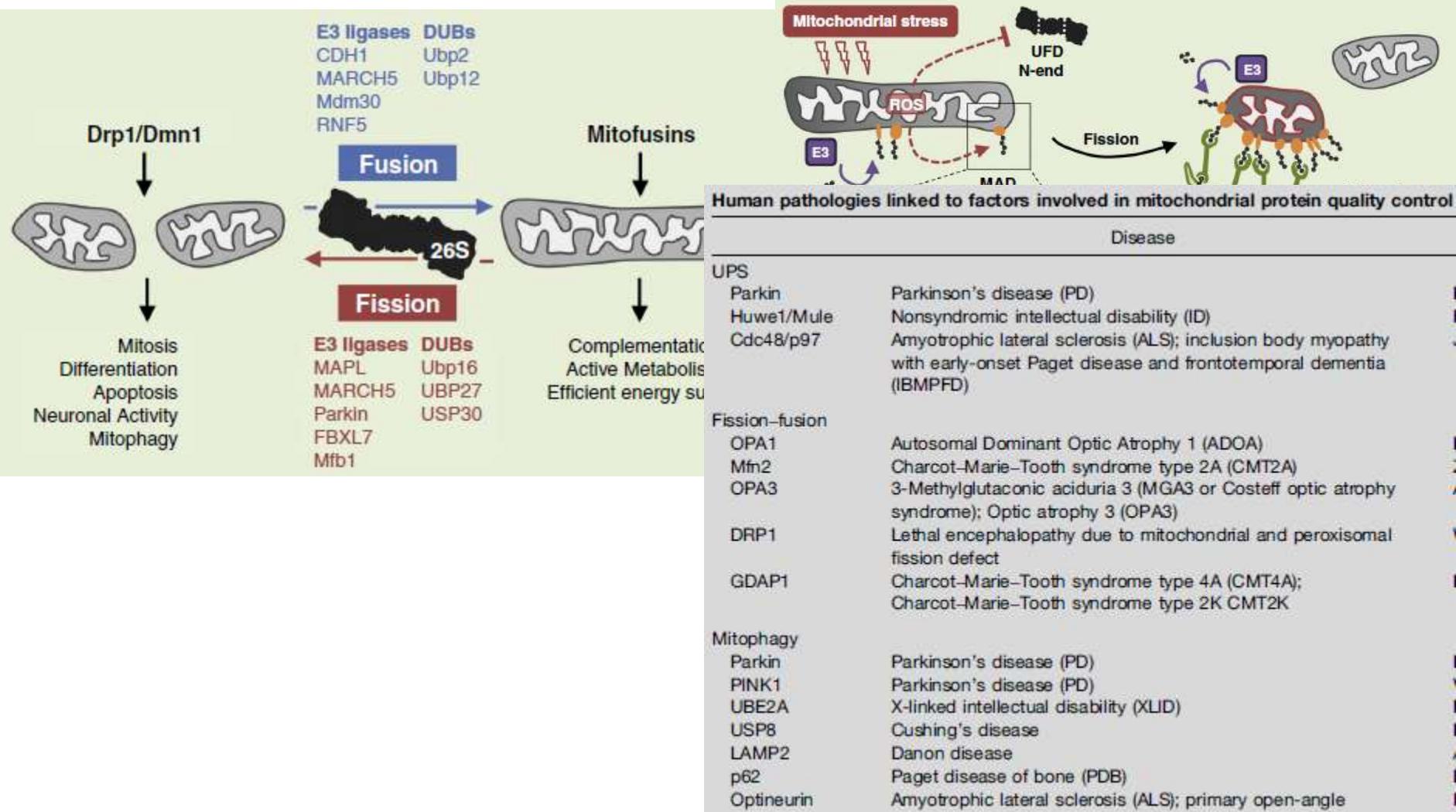
Accepted Date: 13 September 2015

# Autophagy Dysfunction and Consequences

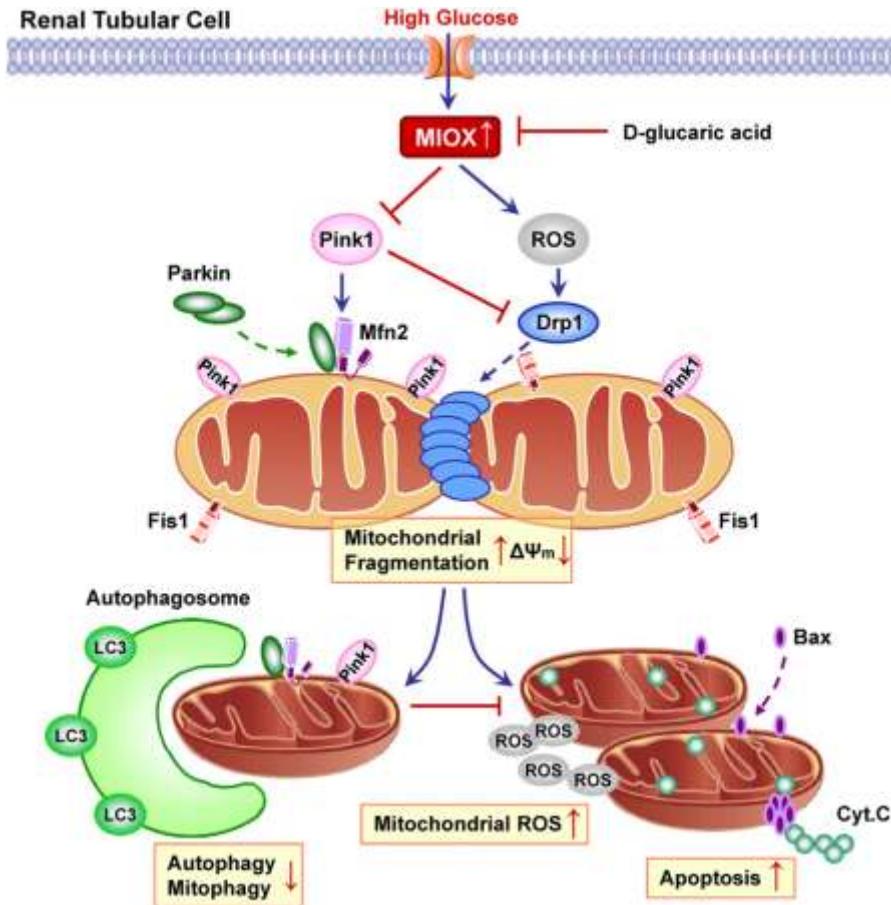
# Autophagy Dysfunction: Consequences



# Mitophagy Dysfunction: Consequences



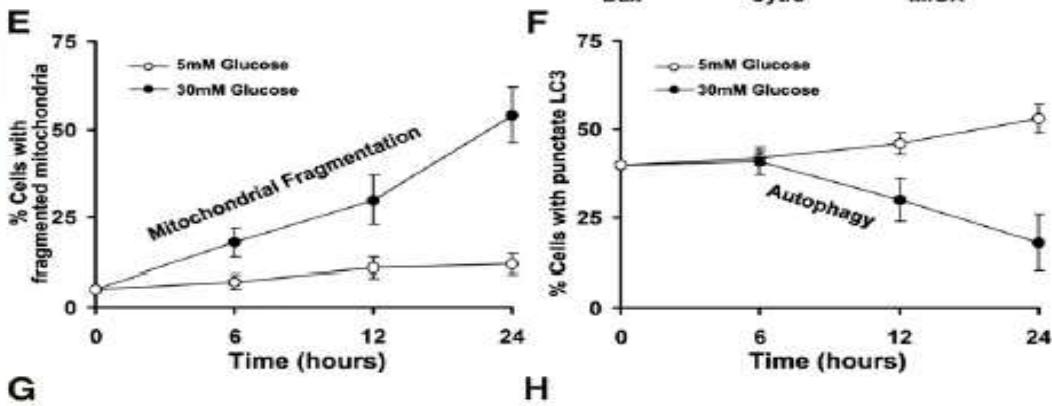
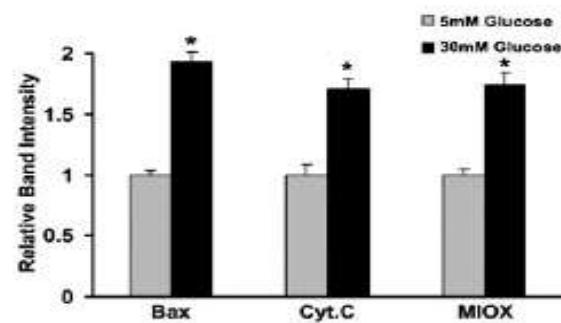
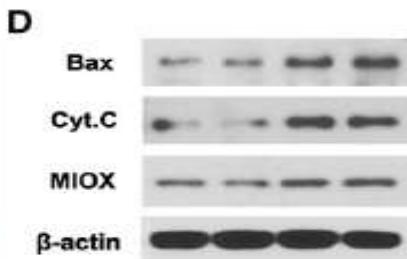
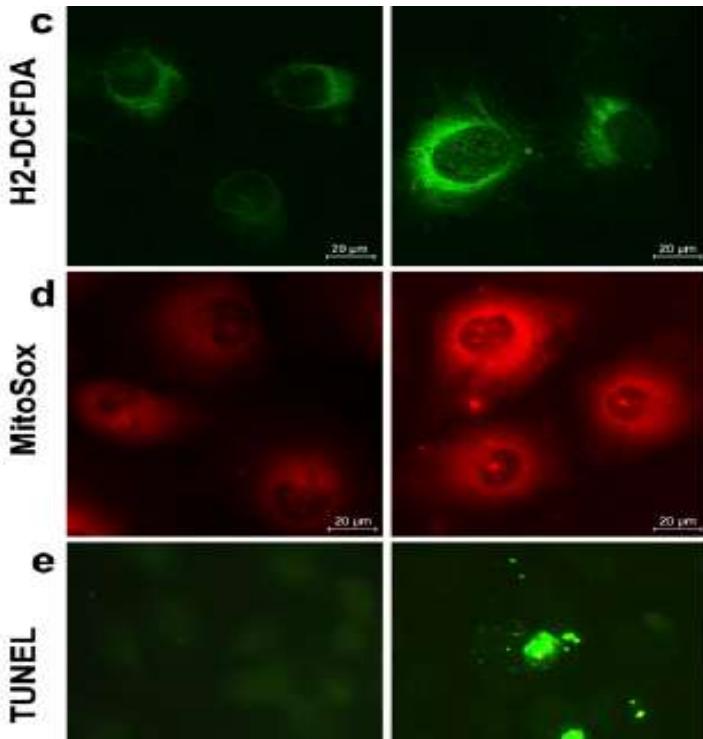
# MIOX Pathway



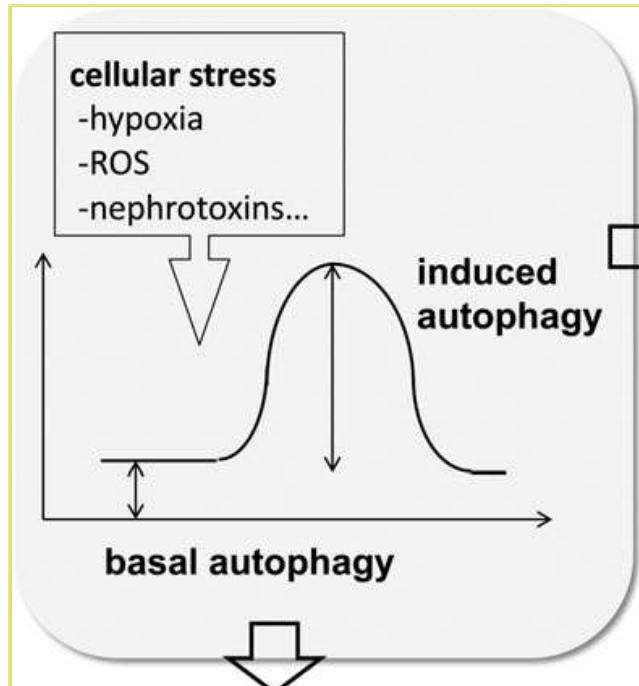
# Mitophagy Dysfunction: MIOX Pathway

BASIC RESEARCH

[www.jasn.org](http://www.jasn.org)



# Autophagy Dysfunction: Kidney Diseases



Autophagy coordinates cellular homeostasis in podocytes and proximal tubules

- clearance of aggregates and damaged mitochondria
- preserving kidney function and slowing aging

**BASIC RESEARCH**

[www.jasn.org](http://www.jasn.org)

# Deficient Autophagy Results in Mitochondrial Dysfunction and FSGS

Takahisa Kawakami,<sup>\*†‡</sup> Ivan G. Gomez,<sup>\*†‡</sup> Shuyu Ren,<sup>\*†§</sup> Kelly Hudkins,<sup>\*</sup> Allie Roach,<sup>\*†§</sup> Charles E. Alpers,<sup>\*</sup> Stuart J. Shankland,<sup>\*</sup> Vivette D. D'Agati,<sup>||</sup> and Jeremy S. Duffield<sup>\*†§</sup>

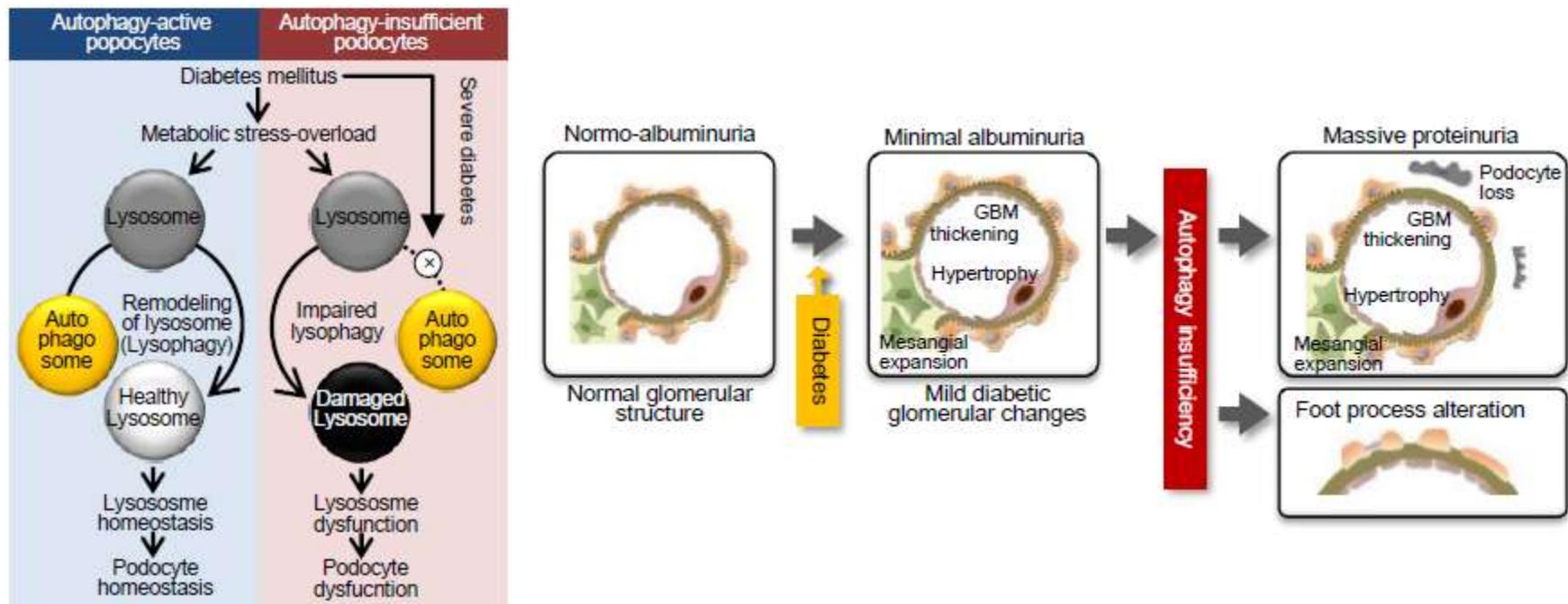
<sup>\*</sup>Division of Nephrology, Departments of Medicine & Pathology, and <sup>†</sup>Institute for Stem Cell & Regenerative Medicine, University of Washington, Seattle, Washington; <sup>‡</sup>Division of Nephrology and Endocrinology, The University of Tokyo, Tokyo, Japan;

<sup>§</sup>Biogen Idec, Inc., Cambridge, Massachusetts; and <sup>||</sup>Department of Pathology, Columbia University, New York, New York

J Am Soc Nephrol 26: 1040–1052, 2015

# Autophagy Dysfunction: Podocytopathy in DN

Impaired podocyte autophagy exacerbates proteinuria in diabetic nephropathy



Diabetes 2015, Accepted

# Endothelial cell and podocyte autophagy synergistically protect from diabetes-induced glomerulosclerosis

Olivia Lenoir,<sup>1,2</sup> Magali Jasiek,<sup>1,2</sup> Carole Hénique,<sup>1,2</sup> Léa Guyonnet,<sup>1,2</sup> Björn Hartleben,<sup>3</sup> Tillmann Bork,<sup>3</sup> Anna Chipont,<sup>1,2</sup> Kathleen Flosseau,<sup>1,2</sup> Imane Bensaada,<sup>1,2</sup> Alain Schmitt,<sup>2,4,5</sup> Jean-Marc Massé,<sup>2,4,5</sup> Michèle Souyri,<sup>6</sup> Tobias B Huber,<sup>3,7</sup> and Pierre-Louis Tharaux<sup>1,2,8,\*</sup>

<sup>1</sup>Paris Cardiovascular Research Center; Institut National de la Santé et de la Recherche Médicale (INSERM); Paris, France; <sup>2</sup>Université Paris Descartes; Sorbonne Paris Cité; Paris, France; <sup>3</sup>Renal Division; University Hospital Freiburg; Freiburg, Germany; <sup>4</sup>CNRS UMR8104; Paris, France; <sup>5</sup>Plateforme de Microscopie Electronique; INSERM U1016; Institut Cochin; Paris, France; <sup>6</sup>CNRS; UMR7622; Paris, France; <sup>7</sup>BLOSS Center for Biological Signaling Studies; Albert-Ludwigs-University; Freiburg, Germany; <sup>8</sup>Nephrology Service; Georges Pompidou European Hospital; Assistance Publique-Hôpitaux de Paris; Paris, France

# ANCA Associated Vascuilitis



**Clinical & Experimental Immunology**  
The Journal of Translational Immunology

British Society for  
**immunology**

Clinical and Experimental Immunology

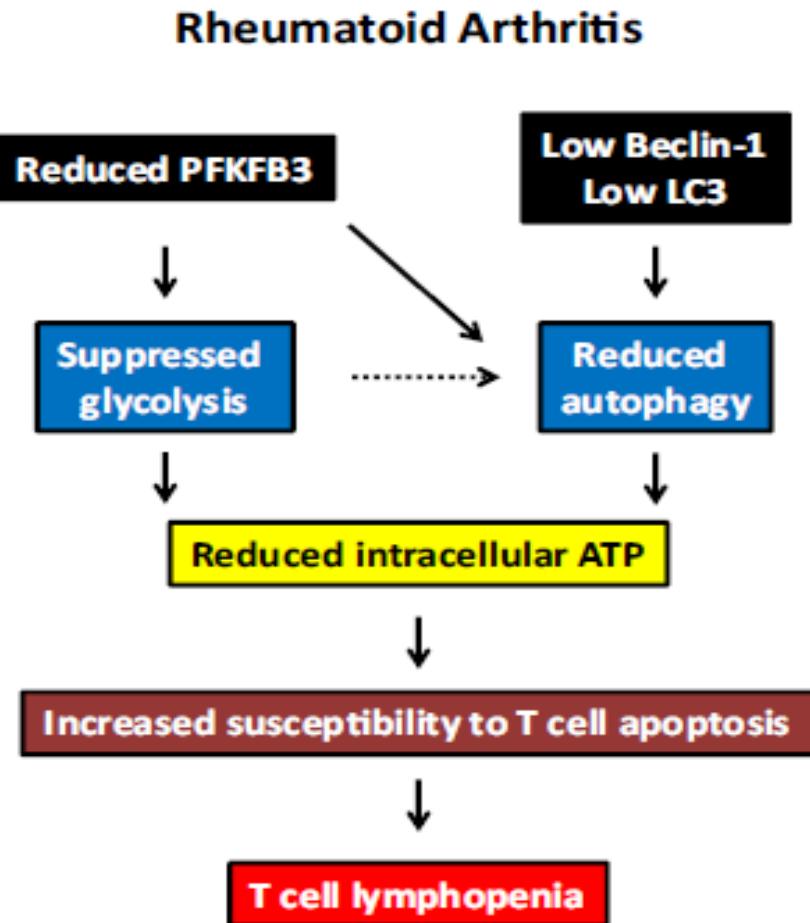
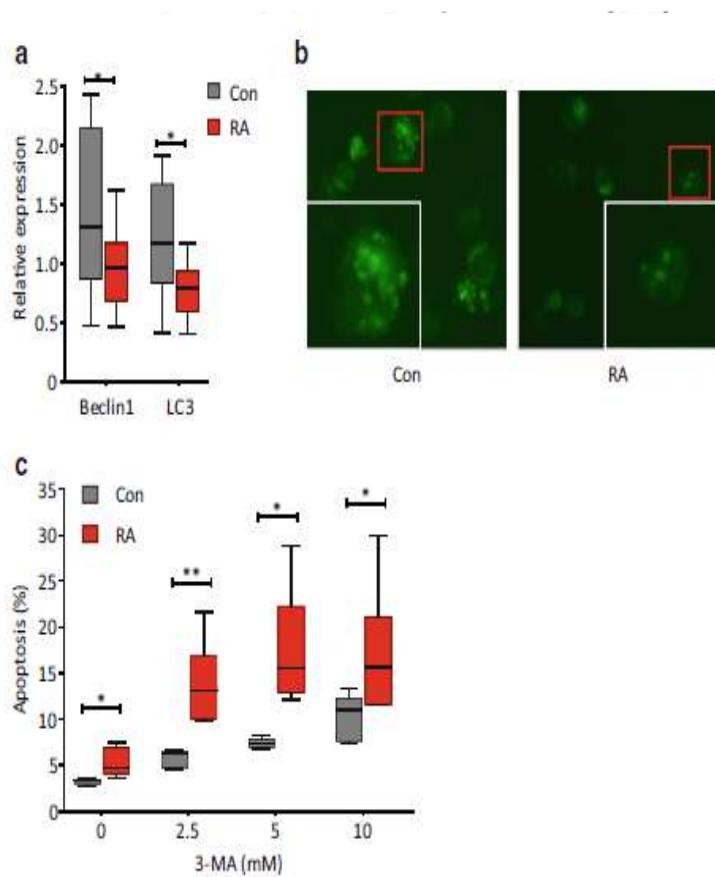
ORIGINAL ARTICLE

doi:10.1111/cei.12589

**Neutrophil extracellular trap formation is associated with autophagy-related signalling in ANCA-associated vasculitis**

Clinical and Experimental Immunology 2015, 180: 408–418

# Autophagy Dysfunction: Autoimmune Disease

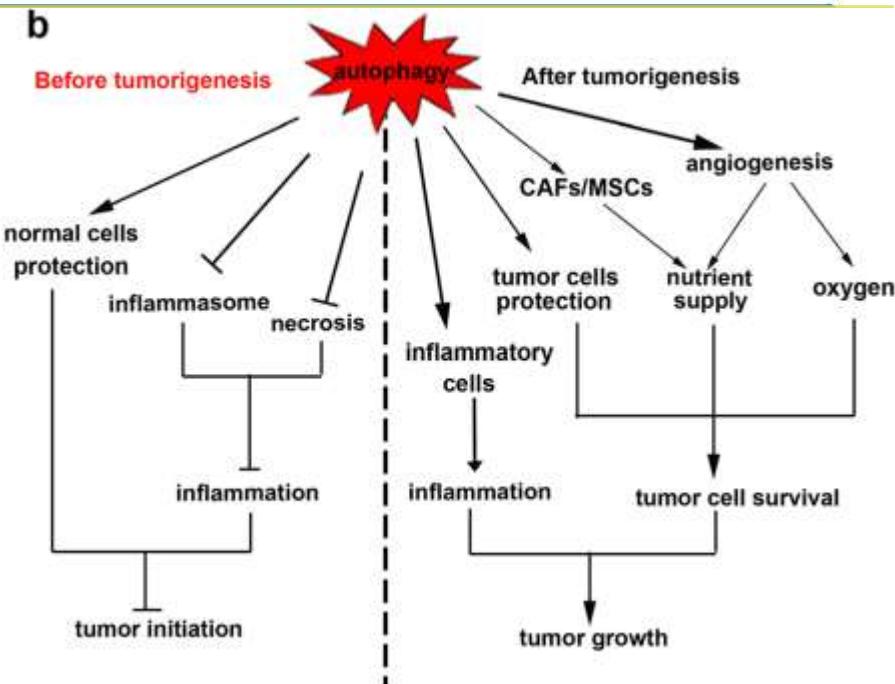
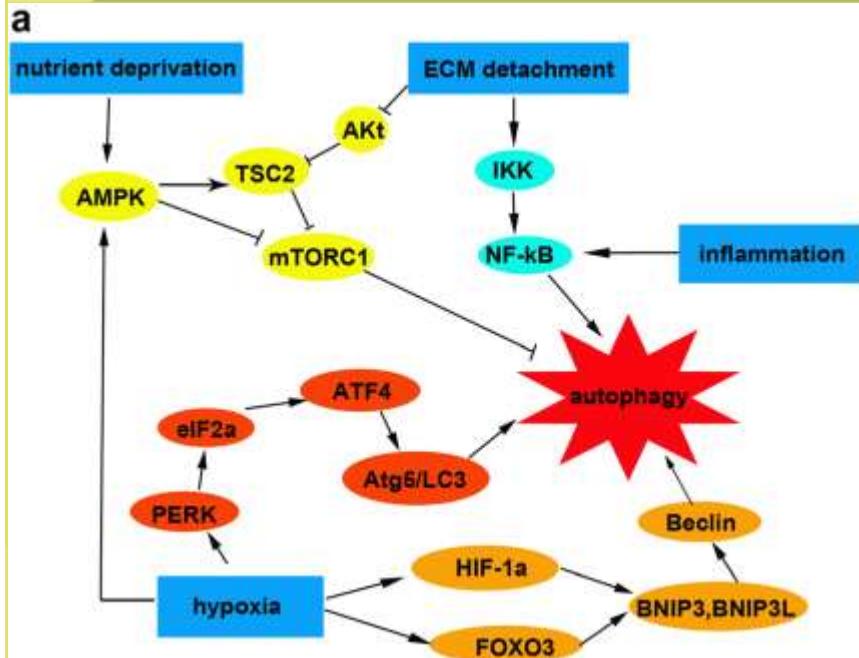


# Autophagy Dysfunction: Oncogenesis

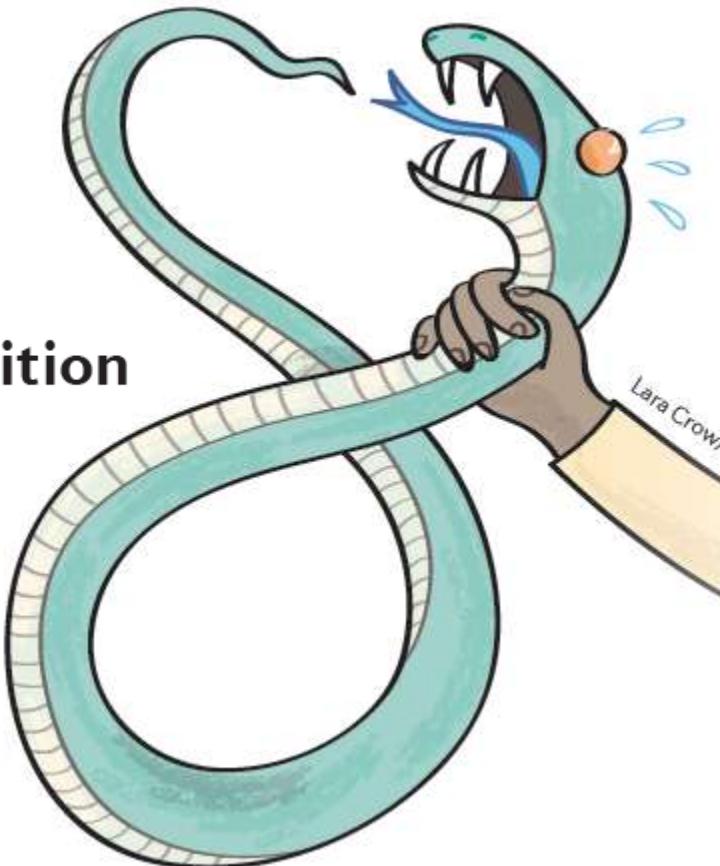
Yang et al. *Cell & Bioscience* (2015) 5:14  
DOI 10.1186/s13578-015-0005-2



*Cell & Bioscience*



# Exploring the anticancer effects of autophagy inhibition



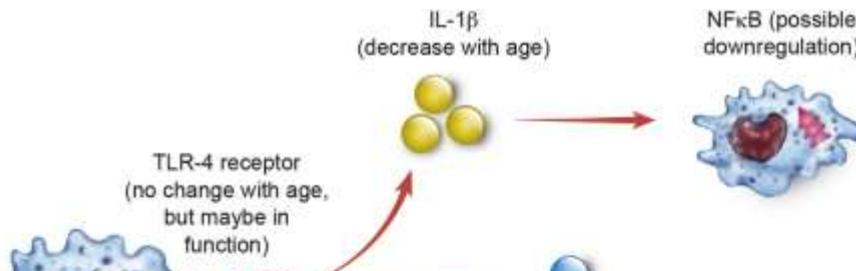
“ these results indicate that the inhibition of autophagy in the intestinal epithelium may suppress the development and progression of tumours ”

**ORIGINAL RESEARCH PAPER** Lévy, J. et al.  
Intestinal inhibition of Atg7 prevents tumour initiation through a microbiome-influenced immune response and suppresses tumour growth.  
*Nat. Cell Biol.* 17, 1062–1073 (2015)

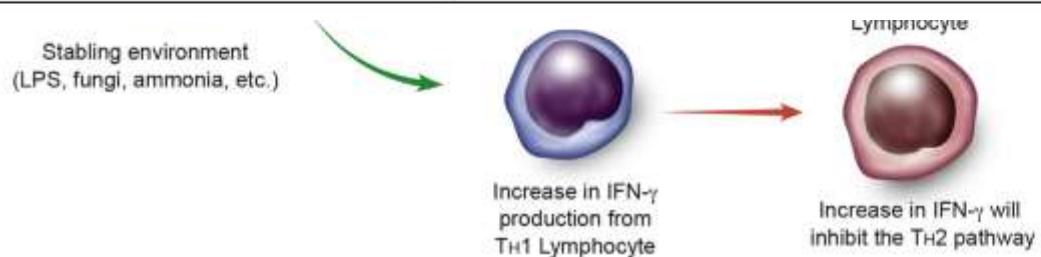
# Autophagy, Immunity and Immunosenescence

# Immunosenescence

## Immunology of the aging equine lung



Cell type	Peripheral blood (human)	Airspace of the lung (human)
Alveolar macrophage	—	↓
Neutrophils	—	↑
Total lymphocytes	—	↑
CD4+ T-cells	↓	
CD8+ T-cells	↓	
CD4+/CD8+ ratio	↓	↑
B-cells	↓	—



# Autophagy Defect: Macrophage Aging



## Research Article

Journal of  
**Innate  
Immunity**

J Innate Immun 2015;7:375–391  
DOI: 10.1159/000370112

Received: June 13, 2014  
Accepted after revision: November 25, 2014  
Published online: March 10, 2015

## Autophagy Controls Acquisition of Aging Features in Macrophages

Amanda J. Stranks<sup>a</sup> Anne Louise Hansen<sup>b</sup> Isabel Panse<sup>a</sup>  
Monika Mortensen<sup>e</sup> David J.P. Ferguson<sup>c</sup> Daniel J. Puleston<sup>a</sup>  
Kevin Shenderov<sup>f</sup> Alexander Scarth Watson<sup>a</sup> Marc Veldhoen<sup>d</sup>  
Kanchan Phadwal<sup>b</sup> Vincenzo Cerundolo<sup>a</sup> Anna Katharina Simon<sup>a, b</sup>

<sup>a</sup>MRC Human Immunology Unit, Weatherall Institute of Molecular Medicine, University of Oxford, <sup>b</sup>BRC Translational Immunology Lab, Experimental Medicine, Nuffield Department of Medicine, and <sup>c</sup>Nuffield Department of Clinical and Laboratory Sciences, John Radcliffe Hospital, Oxford, <sup>d</sup>Babraham Institute, Cambridge, UK; <sup>e</sup>Cell Death and Metabolism, Danish Cancer Society Research Center, Danish Cancer Society, Copenhagen, Denmark; <sup>f</sup>Johns Hopkins University School of Medicine, Baltimore, Md., USA

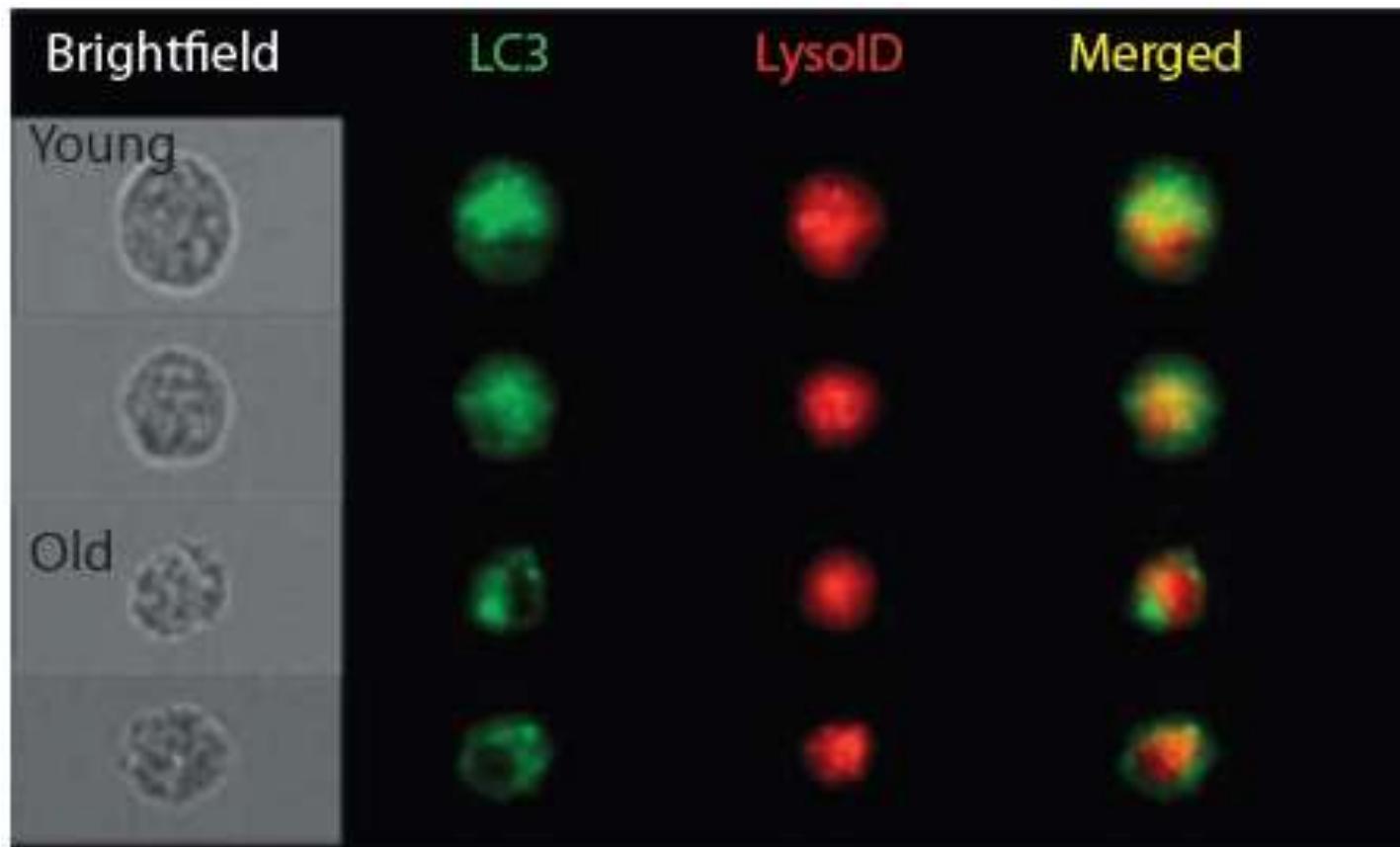
J Innate Immun 2015;7:375–391

# Autophagy Defect: Macrophage Aging

Summary of similarities between aged and Atg7<sup>-/-</sup>-deficient macrophages

Aged macrophages	Macrophage phenotype	Atg7 <sup>-/-</sup> macrophages
✓ (not in peritoneum)	Relatively increased numbers of macrophages, monocytes	✓
✓ (after LPS stimulation)	Increased mitochondria and mROS	✓
✓	Decreased phagocytosis	✓ (phagocytosis of beads)
✓	Decreased NO	✓
✓	Reduced surface marker expression	✓
✓	Reduced antigen presentation (DC)	✓
✓	Increased inflammatory cytokine production	✓
✓	Metabolism alterations	✓

# Autophagy Defect: Macrophage Aging



# Autophagy Defect: Macrophage Aging

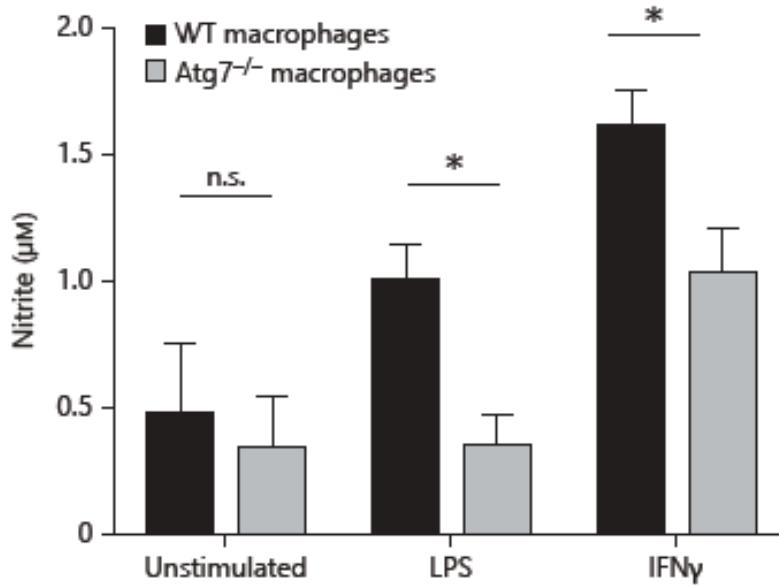
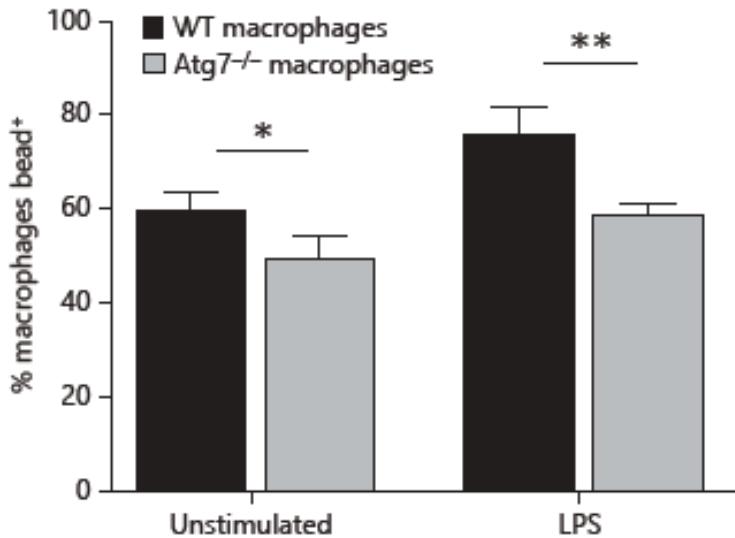


Summary of surface marker expression on autophagy-deficient macrophages relative to WT

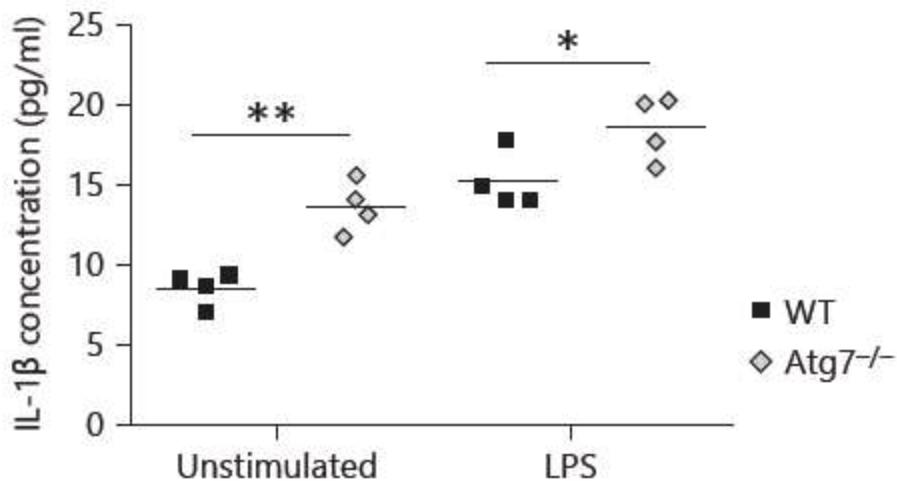
Marker	Atg7 <sup>-/-</sup>
F4/80	±/●
CD11b	●
MHC I	↓
MHC II	↓
CD47	↓
CD48	↓
TLR4	↓
CD86	↓
Mannose receptor	↓
CD13	±
CD14	↓
M-CSFR	↓

The expression of each marker [decreased (↓), similar (●) or variable (±)] was determined by the geometric mean of fluorescence relative to WT macrophages.

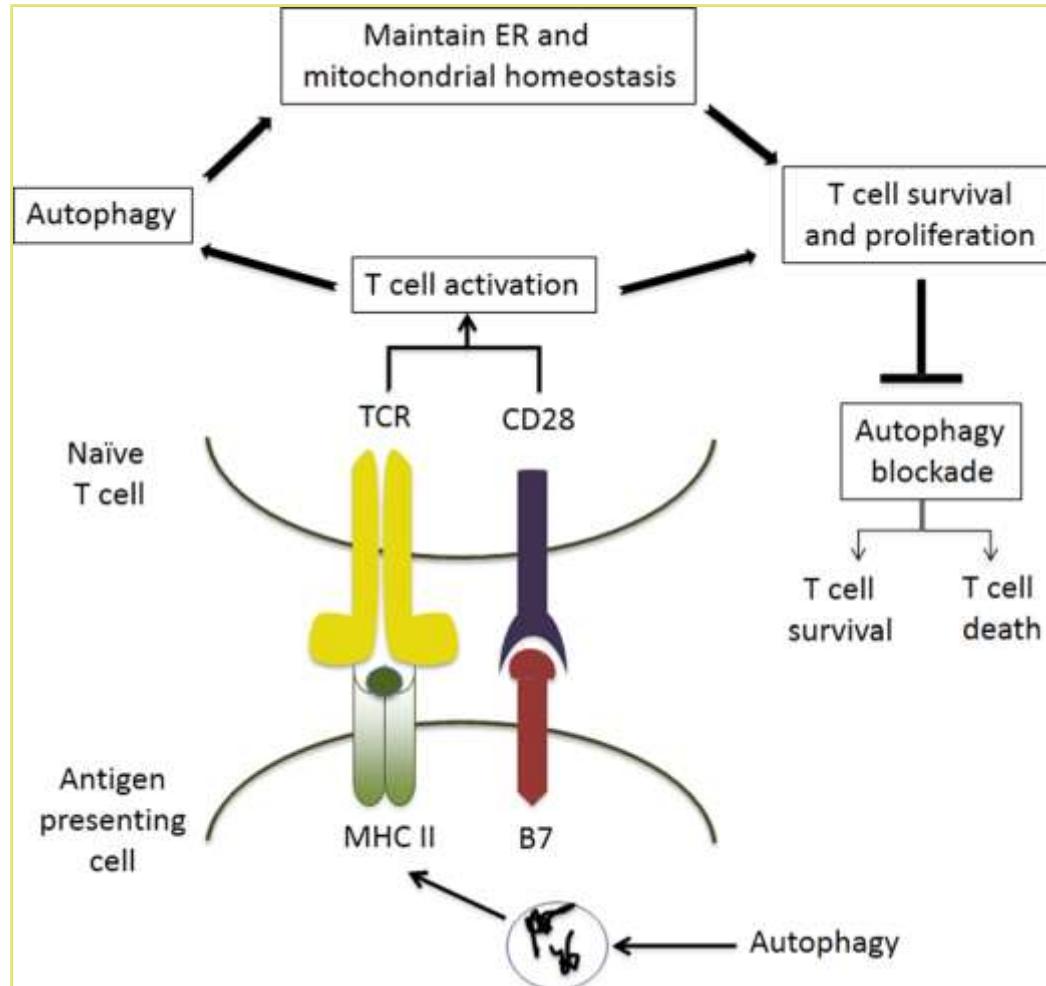
# Autophagy Defect: Macrophage Aging



# Autophagy Defect: Macrophage Aging

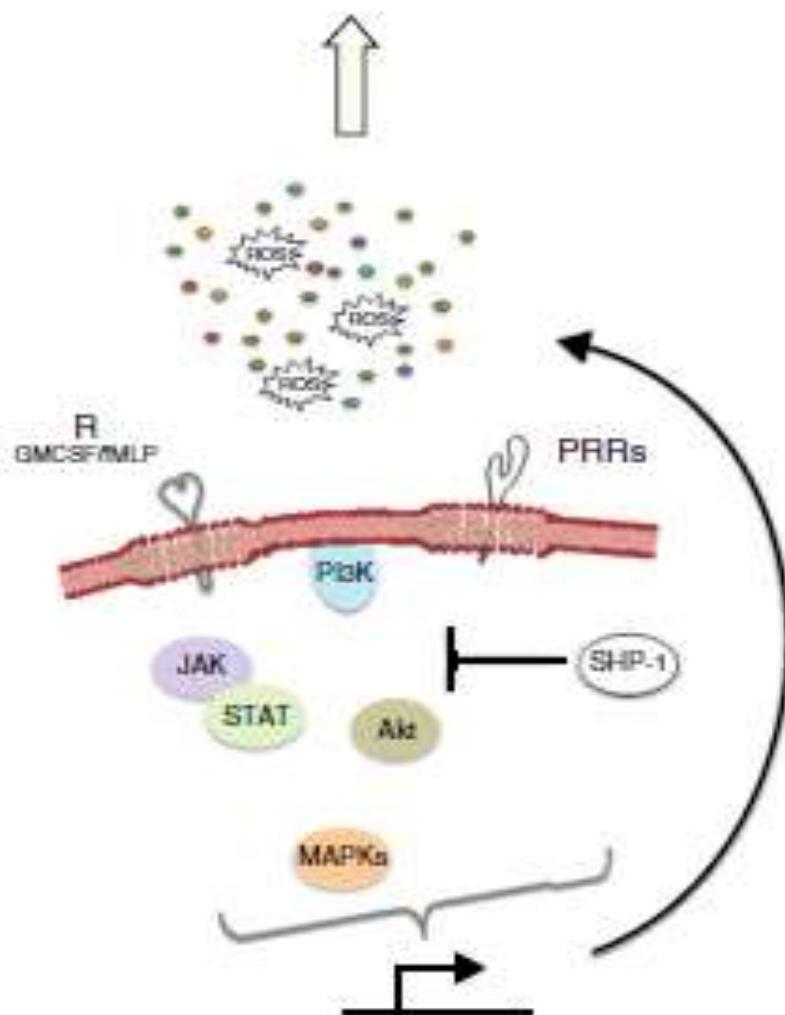


# Autophagy Effects: T Cell



(a)

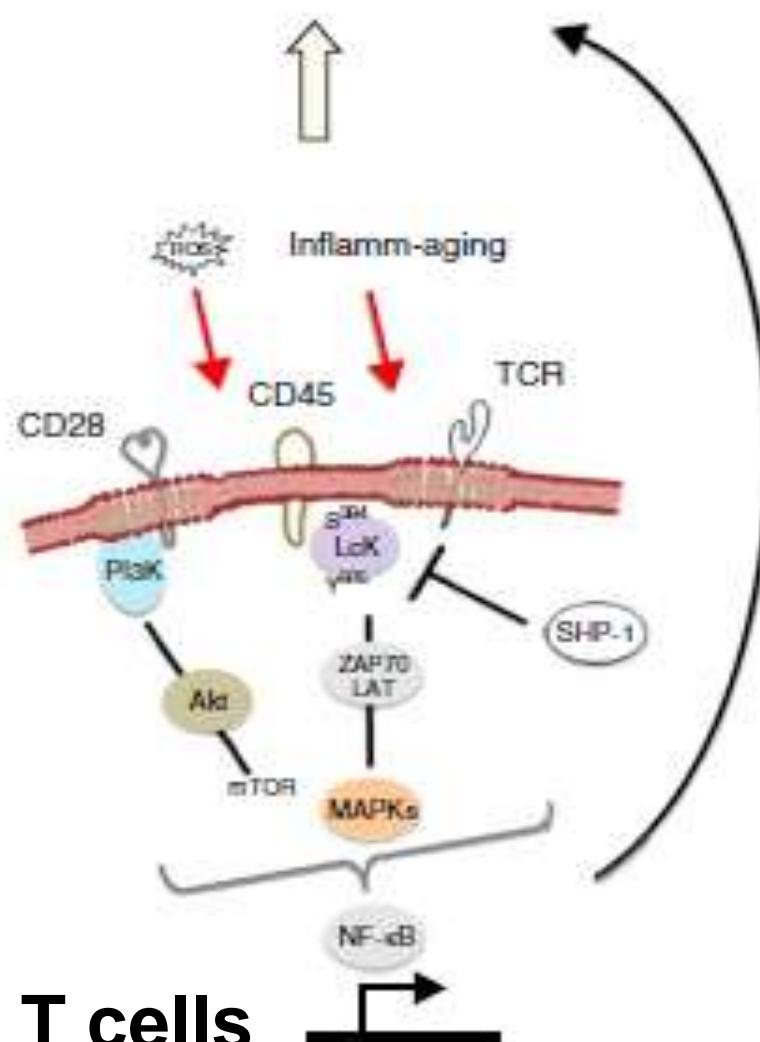
Phagocytosis  
Chemotaxis  
Killing



**neutrophils**

(b)

Proliferation  
Cytokine switch  
Senescence  
Exhaustion



**T cells**

Current Opinion in Immunology

# Autophagy Upregulation: B Cell



**Clinical & Experimental Immunology**  
The Journal of Translational Immunology

British Society for  
**immunology**

Clinical and Experimental Immunology

ORIGINAL ARTICLE

doi:10.1111/cei.12658

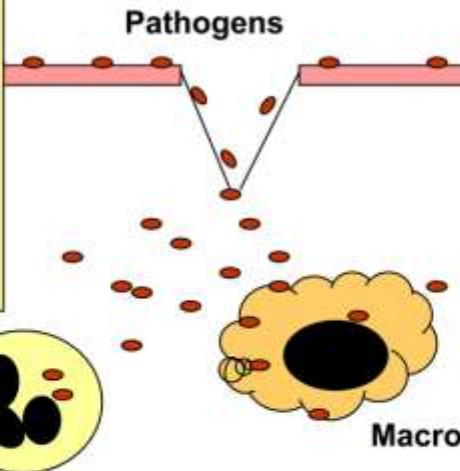
***TRAF3IP3*, a novel autophagy up-regulated gene, is involved in marginal zone B lymphocyte development and survival**

Clinical and Experimental Immunology 2015, 182: 57–68

# Immunosenescence: At A Glance

<u>Neutrophils in the aged</u>	
Preserved	<ul style="list-style-type: none"><li>Number (in healthy elderly)</li><li>Adherence</li><li>TLR expression</li></ul>
Reduced	<ul style="list-style-type: none"><li>Chemotaxis</li><li>Phagocytosis</li><li>SuperOxide production</li><li>Molecules recruitment into lipid raft</li><li>Signal transduction</li><li>Apoptosis</li></ul>

Neutrophils



<u>Macrophages in the aged</u>	
Preserved	<ul style="list-style-type: none"><li>Number (altered subsets)</li></ul>
Reduced	<ul style="list-style-type: none"><li>Chemotaxis</li><li>Phagocytosis</li><li>SuperOxide production</li><li>Signal transduction</li><li>Apoptosis</li><li>TLR expression and function</li><li>MHC class II expression</li><li>Cytokine production</li></ul>
Increased	<ul style="list-style-type: none"><li>PGE2 production</li></ul>

Macrophages



Dendritic cells

<u>Dendritic cells in the aged</u>	
pDC:	<ul style="list-style-type: none"><li>Decreased IFN-I/III production</li><li>Decreased antigen presentation</li></ul>
mDC:	<ul style="list-style-type: none"><li>Decreased TLR-mediated signalling</li><li>Decreased antigen presentation</li><li>Decreased chemotaxis and endocytosis</li></ul>

# Immune Functions of Autophagy



## Immune functions of autophagy

Innate immunity	Adaptive immunity
Regulation of inflammatory cytokine production	Thymic selection
Regulation of type I interferon production	Antigen presentation
Regulation of inflammatory transcriptional response	Apoptotic bodies clearance
Apoptotic bodies clearance	Lymphocytes homeostasis
Pathogen degradation	–
Phagosome maturation	–

## Innate immunity

- Pathogen degradation;
- Antimicrobial peptide/protein;
- Cytoprotection against toxins;
- Apoptotic corpse clearance;
- Regulation of cytokine production;
- Inflammatory transcription regulation;
- Oxidative stress;
- Inflammasome activation
- Pattern recognition receptors (PPRs):
  - PAMP (pathogen-associated molecular patterns): Toll-like receptors (TLR); Nod-like receptors (NLR); RIG-I-like receptors (RLR);
  - DAMP (danger-associated molecular patterns): self-DNA-containing complexes; HMGB1 (high mobility group box1), IL-1 $\beta$ ;
  - SLRs (sequestosome ( SQSTM1/p62)-like receptors): SQSTM1, NBR1, CALCOCO2/NDP52;
  - IRG (Immunity-related GTPases): IRGM

## Adaptive immunity

- Thymic selection;
- T cell maturation;
- T cell polarization;

## Autophagy

- T and B Lymphocyte homeostasis;
- Apoptotic corpse clearance;
- Antigen processing & presentation;
- Antibody response

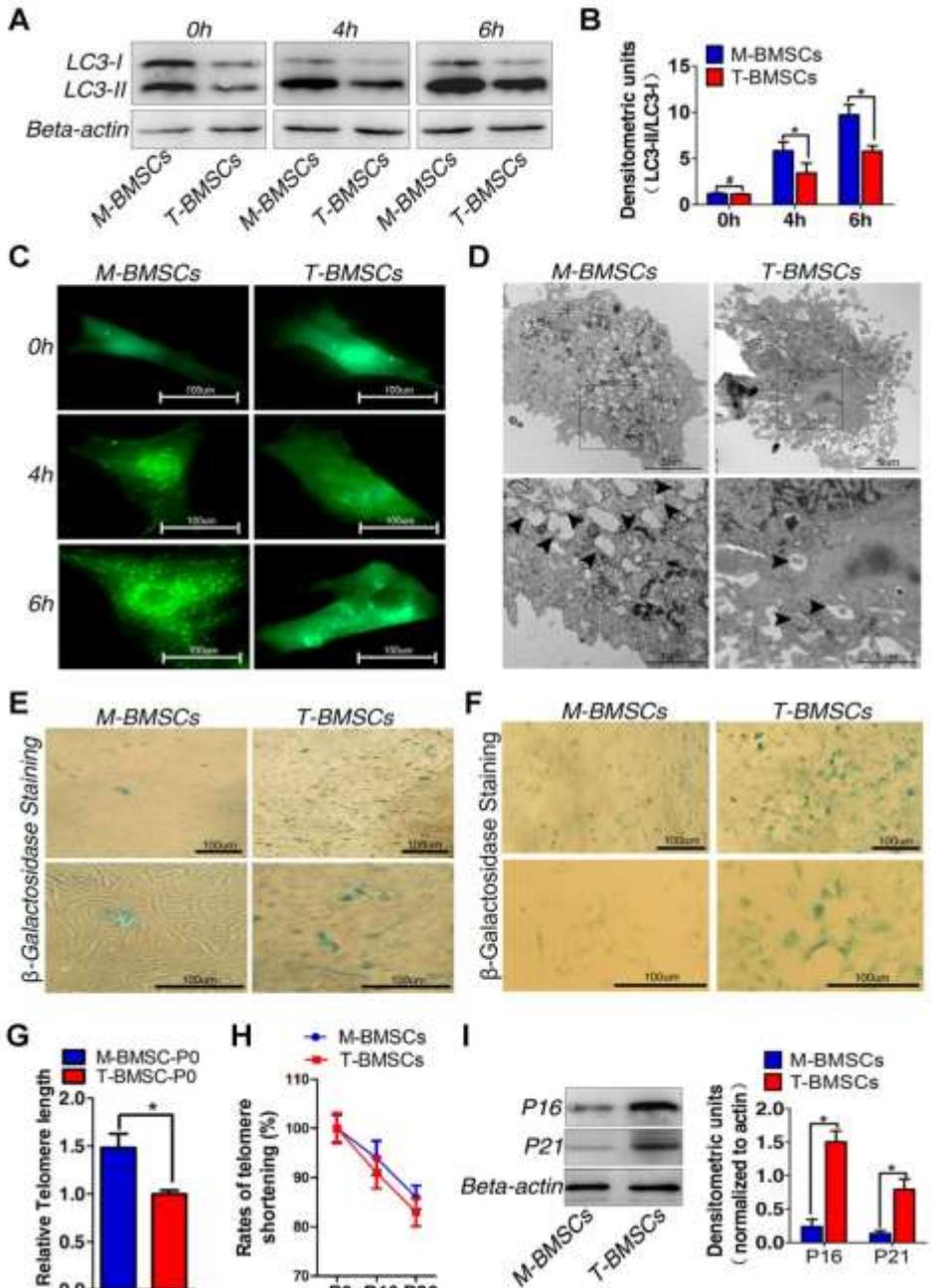
Protection

Balanced responses

Imbalanced responses

Diseasome

# Auto capabili science in T-BMSCs



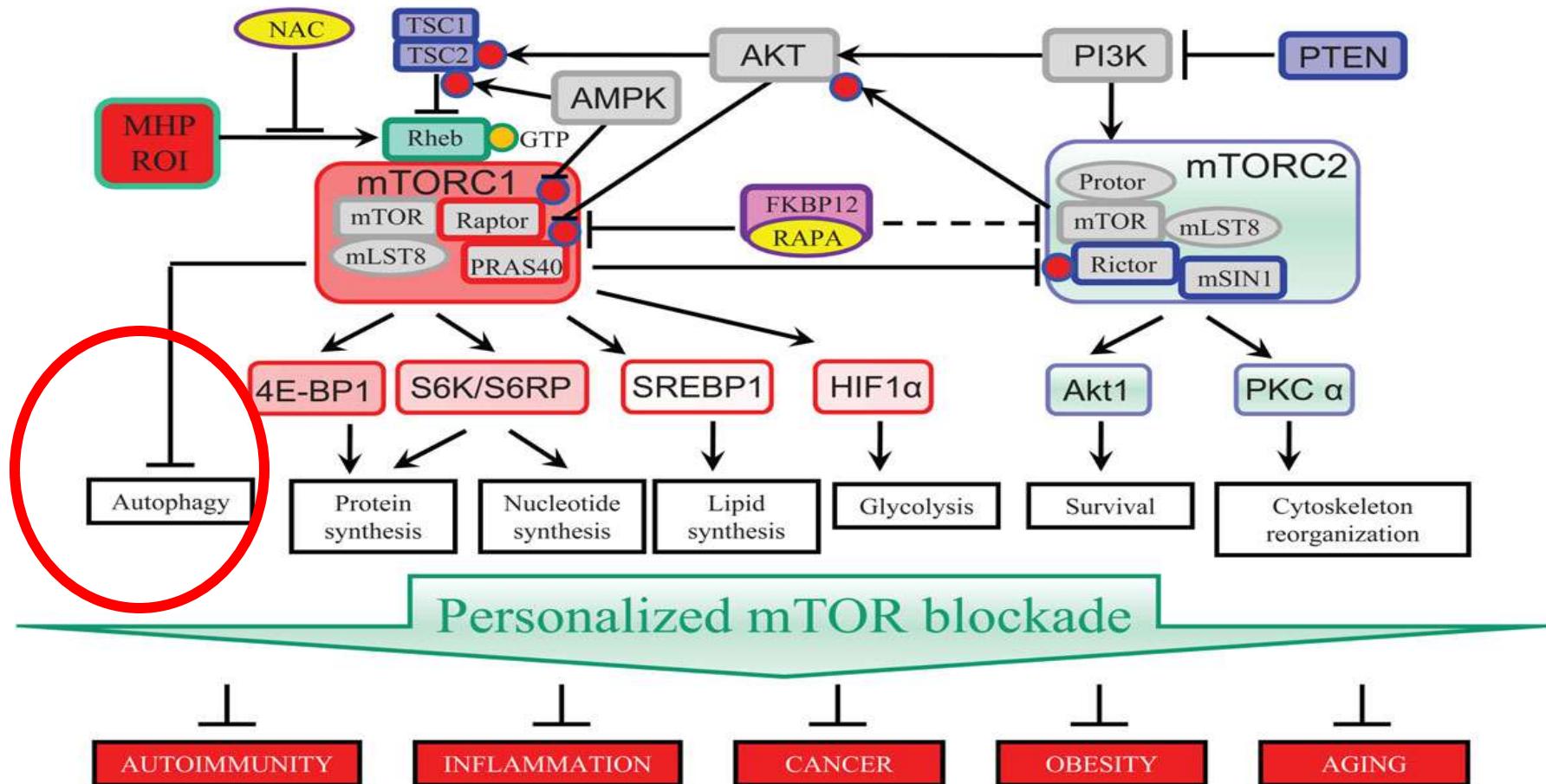


Urology and Nephrology  
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# Autophagy and mTOR

# mTOR In Focus



# mTOR In Focus



## Pharmacological blockade of mTOR pathway activation

Drug	Molecular target	Mechanism of action	Pathway blockade
Rapamycin	FKBP12	Allosteric	mTORC1
Everolimus	FKBP12	Allosteric	mTORC1
Temsirolimus	FKBP12	Allosteric	mTORC1
Torin1	mTOR kinase domain	ATP competitive	mTORC1/mTORC2
AZD8055	mTOR kinase domain	ATP competitive	mTORC1/mTORC2
INK128	mTOR kinase domain	ATP competitive	mTORC1/mTORC2
BGT226	PI3K/mTOR kinase	ATP competitive	PI3K/mTORC1/mTORC2
BAL	Disulfide bonds	H donor	Raptor/mTORC1
NAC	GSH	Antioxidant	mTORC1
Metformin	ETC complex I	Antioxidant	mTORC1

---

CLINICAL RESEARCH PAPER

Autophagy 10:8, 1391–1402; August 2014; © 2014 Landes Bioscience

# Combined MTOR and autophagy inhibition

## Phase I trial of hydroxychloroquine and temsirolimus in patients with advanced solid tumors and melanoma

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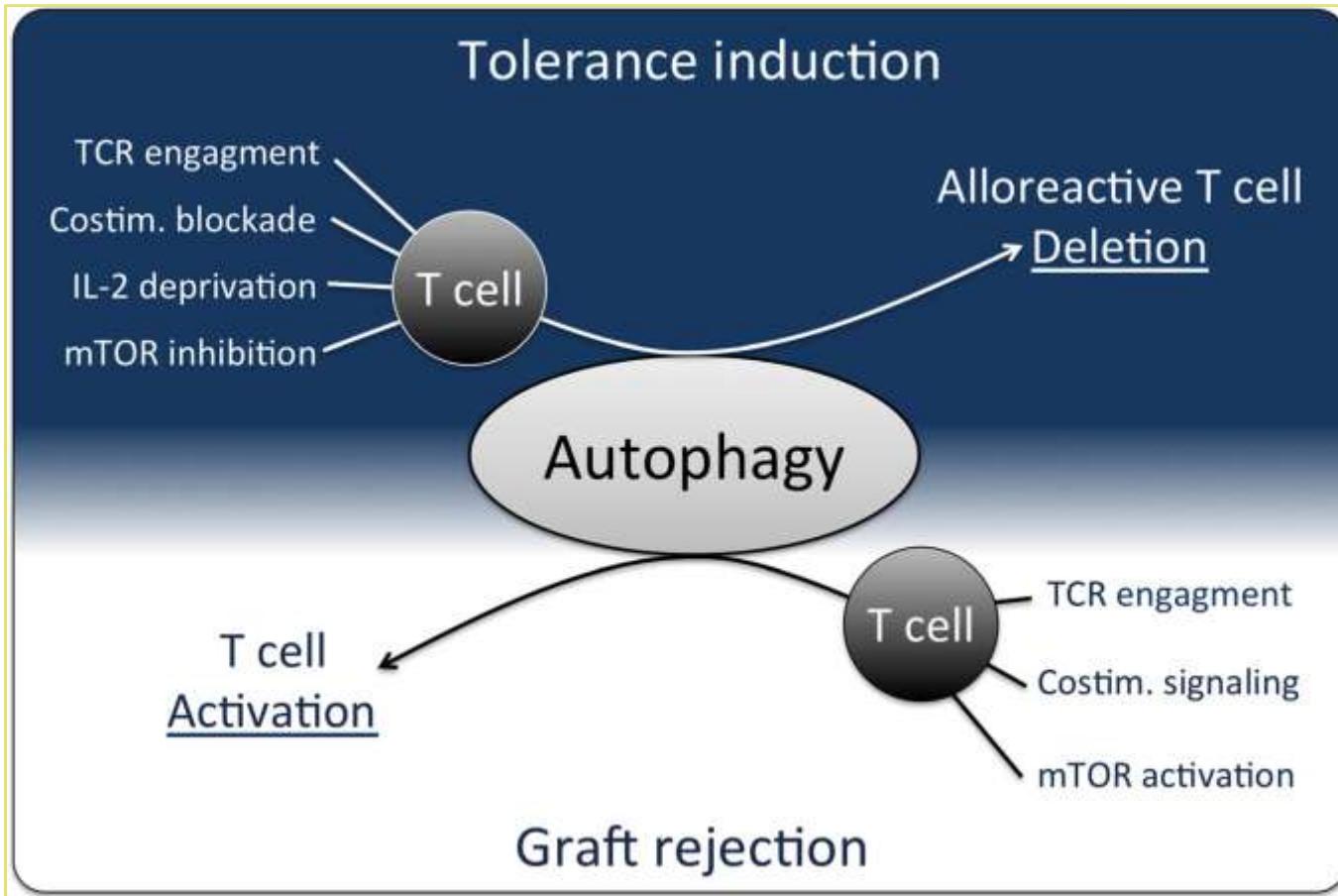
## siRNA based silencing of Mammalian target of rapamycin (mTOR): Implications for Tauopath



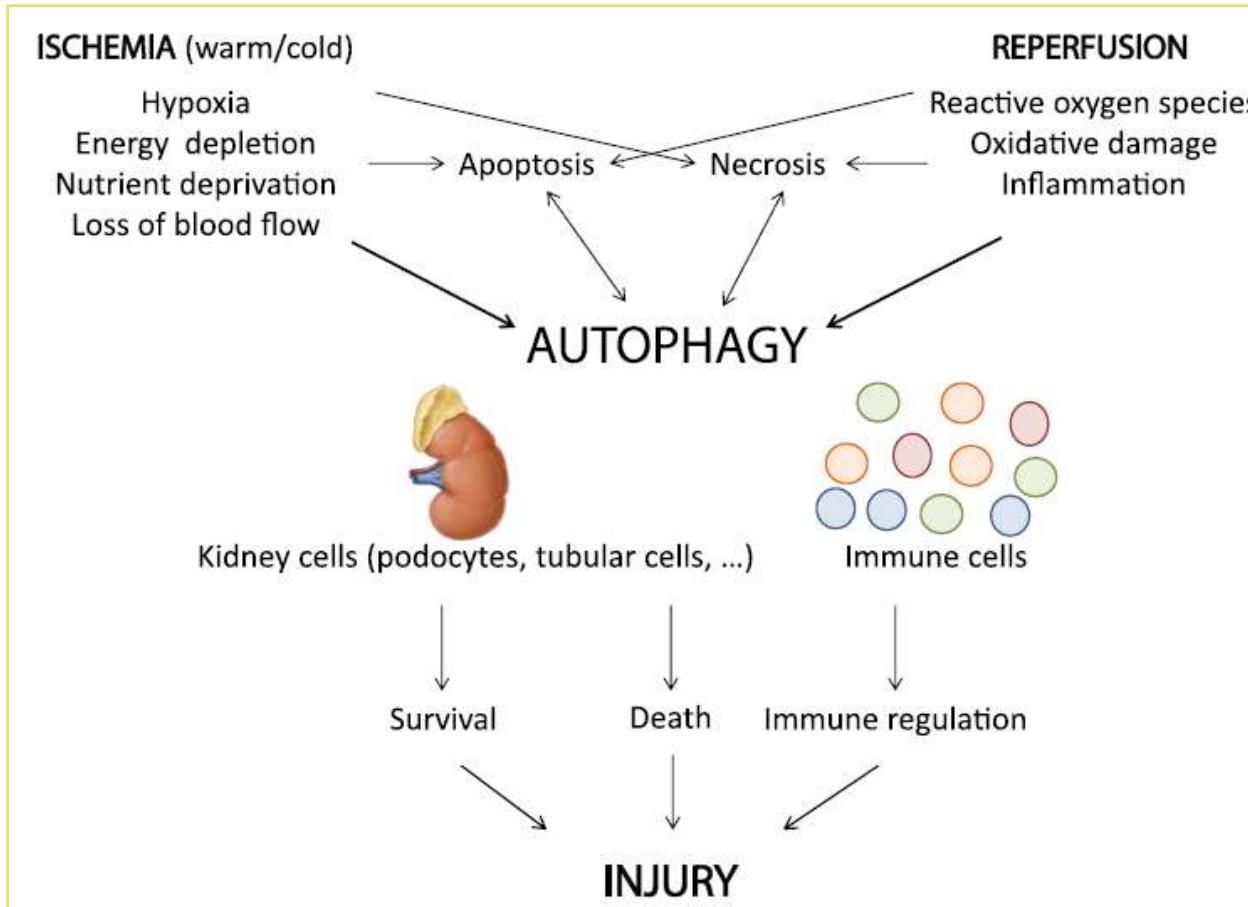
**Dr. Mohamed Salama, Mansoura University**

# Autophagy and Transplantation

# Autophagy: Transplant Immunity



# Autophagy Dysfunction: IRI



ORIGINAL ARTICLE

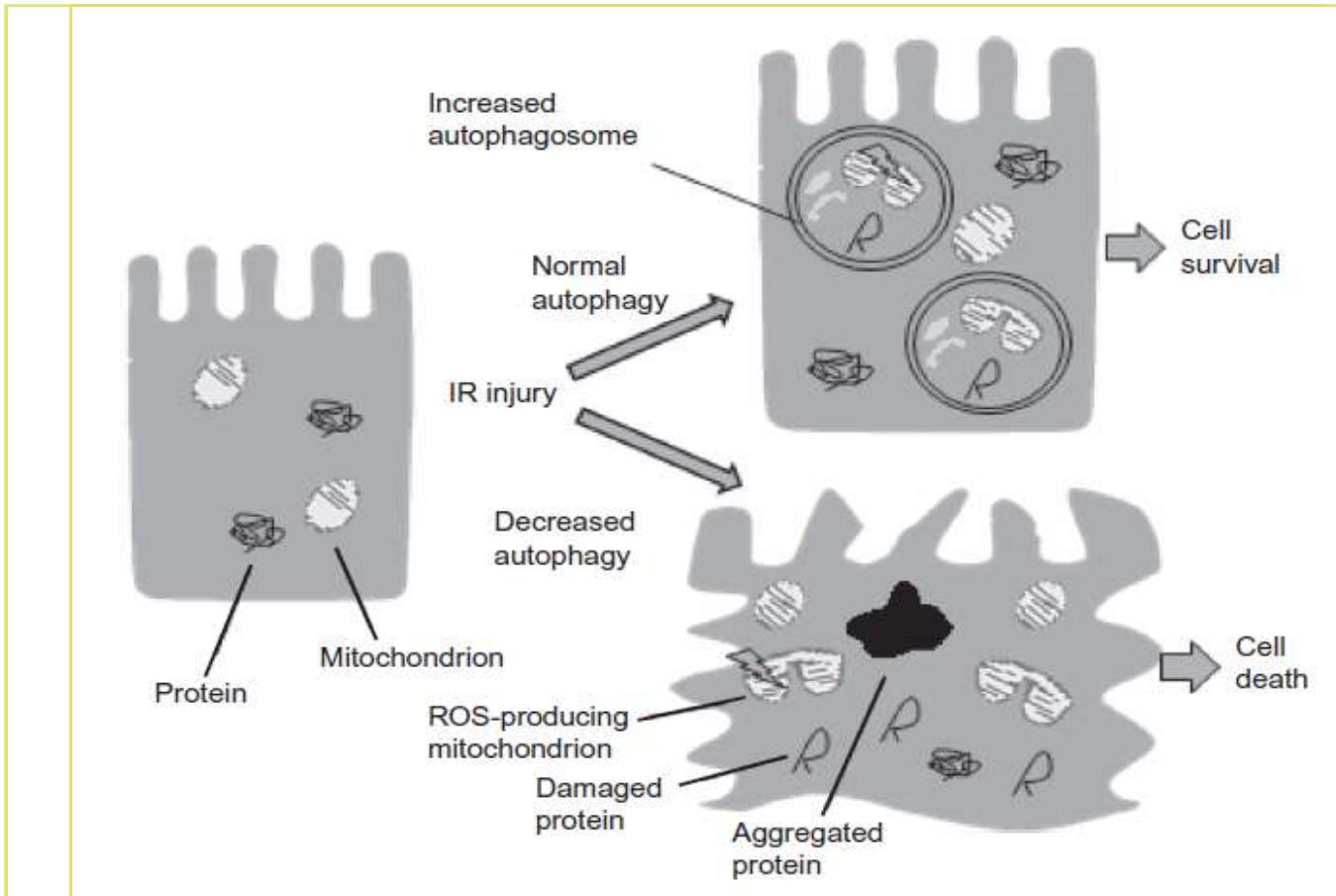
## **Inhibition of autophagy increases apoptosis during re-warming after cold storage in renal tubular epithelial cells**

Swati Jain,<sup>1</sup> Daniel Keys,<sup>1</sup> Trevor Nydam,<sup>1</sup> Robert J. Plenter,<sup>1</sup> Charles L. Edelstein<sup>1,2</sup> and Alkesh Jani<sup>1,2</sup>

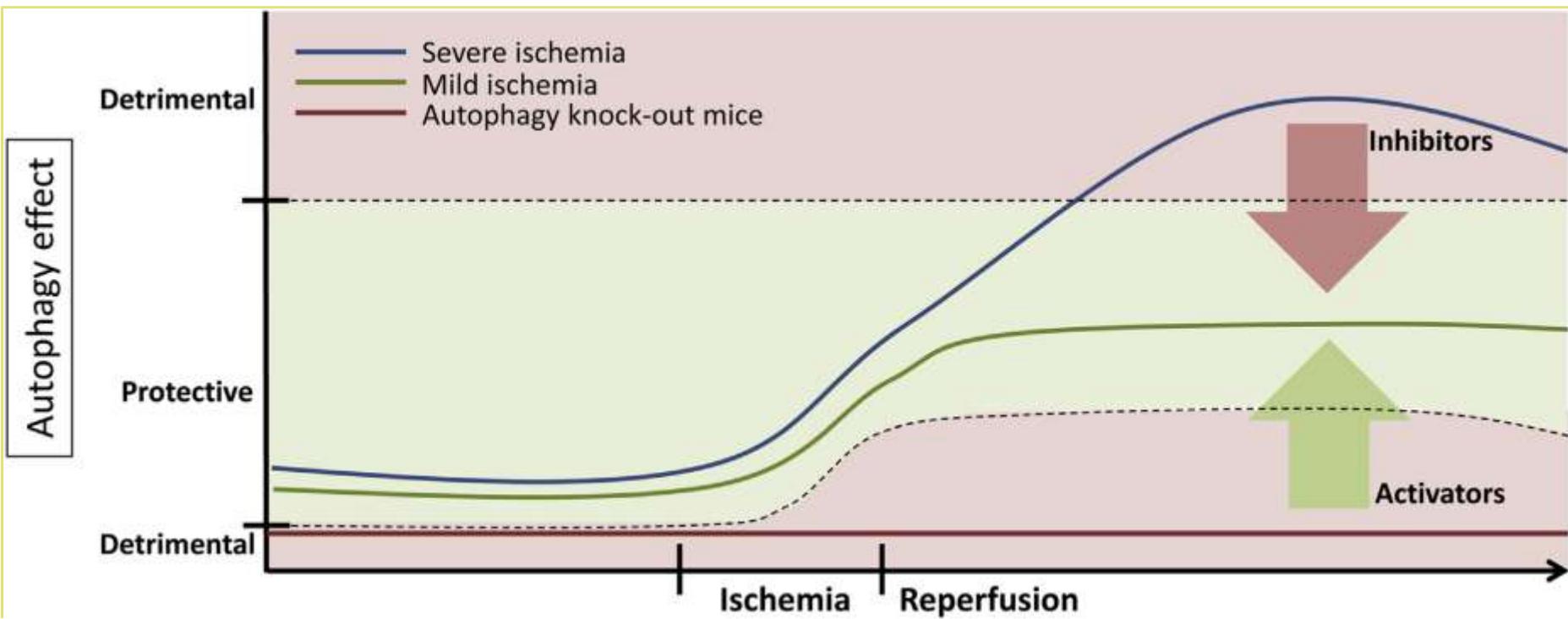
<sup>1</sup> University of Colorado, Denver, CO, USA

<sup>2</sup> Denver Veterans Affairs Medical Center, Denver, CO, USA

# Autophagy: IRI and Immunosuppression

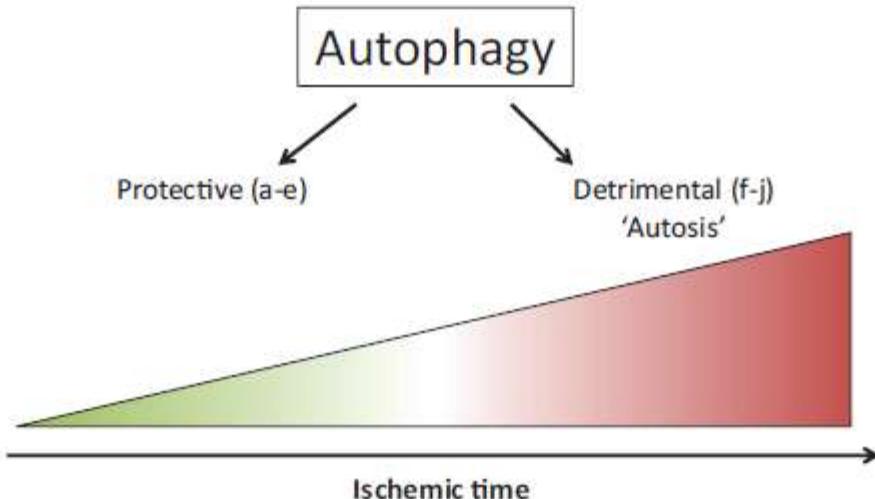


# Autophagy Dysfunction: IRI



# Autophagy: IRI

## Autophagy in Renal Ischemia-Reperfusion Injury: Friend or Foe?



Publication	Ischemia	Publication	Ischemia
(a) Liu S Autophagy 2012; 8(5)	25 min	(f) Nakagawa S Eur J Pharmacol 2012; 696(1-3)	40 min
(b) Jiang M Kidney Int 2012; 82(12)	25 min	(g) Chien CT Transplantation 2007; 84(9)	45 min
(c) Jiang M Am J Pathol 2010; 176(3)	30 min	(h) Isailo Y Transplant Proc 2009; 41(1)	45 min
(d) Kimura T J Am Soc Nephrol 2011; 22(5)	40 min	(i) Yeh CH Life Sci 2010; 86(3-4)	45 min
(e) Lempainen J Acta Physiol 2013; 208(4)	40 min	(j) Wu HH J Biomed Sci 2009; 16(1)	60 min

# Autophagy: IRI



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Experimental Cell Research ■ (■■■) ■■■-■■■



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Experimental Cell Research

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## Research Article

MiR-21 inhibits autophagy by targeting Rab11a in renal ischemia/reperfusion

Xiujuan Liu <sup>a,\*</sup>, Quan Hong <sup>b</sup>, Zhen Wang <sup>a</sup>, Yanyan Yu <sup>a</sup>, Xin Zou <sup>a</sup>, Lihong Xu <sup>a</sup>

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<sup>b</sup> Department of Nephrology, Chinese PLA General Hospital, Chinese PLA Institute of Nephrology, State Key Laboratory of Kidney Diseases, National Clinical Research Center for Kidney Diseases, Beijing 100853, China

Ham et al. *Stem Cell Research & Therapy* (2015) 6:147  
DOI 10.1186/s13287-015-0134-x



RESEARCH

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# let-7b suppresses apoptosis and autophagy of human mesenchymal stem cells transplanted into ischemia/reperfusion injured heart by targeting caspase-3

Onju Ham<sup>1†</sup>, Se-Yeon Lee<sup>1†</sup>, Chang Youn Lee<sup>2†</sup>, Jun-Hee Park<sup>2</sup>, Jiyun Lee<sup>3</sup>, Hyang-Hee Seo<sup>3</sup>, Min-Ji Cha<sup>1,4</sup>, Eunhyun Choi<sup>1,4</sup>, Soonhag Kim<sup>1,4</sup> and Ki-Chul Hwang<sup>1,4\*</sup>

# Autophagy:

## Effect of Immunosuppressants

Rapamycin	Stimulation in vitro and in vivo	Methylprednisolone	Stimulation in vivo in mouse osteocytes
Tacrolimus	Stimulation in vivo in mouse brain	Azathioprine	Inhibition of autophagy-dependent cell death in vivo in rat spinal cord
Mycophenolate mofetil	Stimulation of chaperone-mediated autophagy in vitro in hepatocytes	Cyclosporine A	Stimulation in vitro in hepatoblastoma cells
Dexamethasone	Stimulation in vitro in thymoma lymphocytes Stimulation of autophagy-dependent cell death in vitro in lymphoid leukemia cells Stimulation in vivo in rat soleus muscle Stimulation in vitro in primary osteocytes Stimulation in vitro in chondrocytes		Stimulation in vivo in rat acinar cells Stimulation in vitro in rat pituitary cells Inhibition of autophagic flux in vivo in mouse kidney Stimulation of autophagy-dependent cell death in vitro in glioma cells Stimulation of autophagy-dependent cell death in vitro in rat pituitary cells

# Rapamycin Causes Upregulation of Autophagy and Impairs Islets Function Both *In Vitro* and *In Vivo*

M. Tanemura<sup>a,\*</sup>, Y. Ohmura<sup>a</sup>, T. Deguchi<sup>a</sup>,  
T. Machida<sup>a</sup>, R. Tsukamoto<sup>a</sup>, H. Wada<sup>a</sup>,  
S. Kobayashi<sup>a</sup>, S. Marubashi<sup>a</sup>, H. Eguchi<sup>a</sup>,  
T. Ito<sup>b</sup>, H. Nagano<sup>a</sup>, M. Mori<sup>a</sup> and Y. Doki<sup>a</sup>

Departments of <sup>a</sup>Gastroenterological Surgery and  
<sup>b</sup>Complementary and Alternative Medicine, Osaka  
University Graduate School of Medicine, Osaka, Japan  
*\*Corresponding author: Masahiro Tanemura,*  
*mtanemura@gesurg.med.osaka-u.ac.jp*

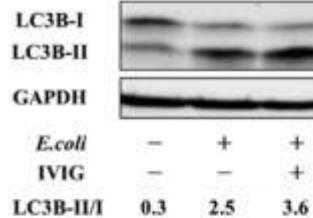
*American Journal of Transplantation 2012; 12: 102–114*



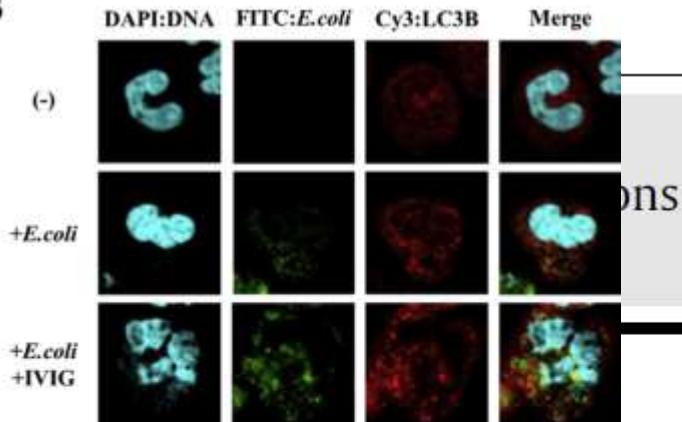
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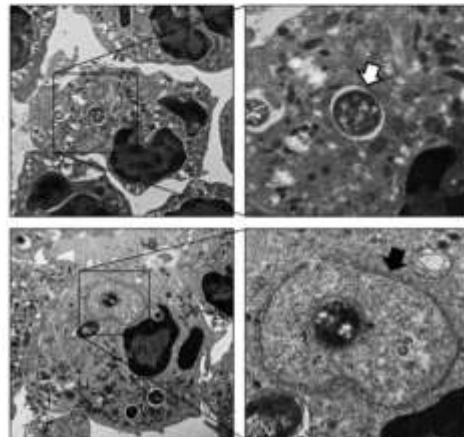
B



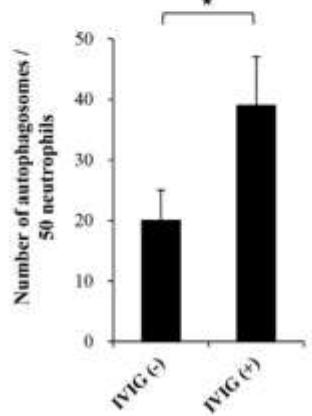
## Intravenous immunoglobulin enhances autophagy of neutrophils against *E. coli* in patients against

Hidemasa Matsuo <sup>a</sup>,  
Takeshi Higuchi <sup>b</sup>, Shuji Akifumi Takaori-Kobayashi <sup>c</sup>

IVIG (-)



IVIG (+)

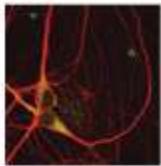


<sup>a</sup> Department of Human Health Sciences, Kyushu University, Fukuoka, Japan

<sup>b</sup> Department of Clinical Laboratory, Kyushu University, Fukuoka, Japan

<sup>c</sup> Department of Infection Control, Kyushu University, Fukuoka, Japan

<sup>d</sup> Department of Hematology and Internal Medicine, Kyushu University, Fukuoka, Japan

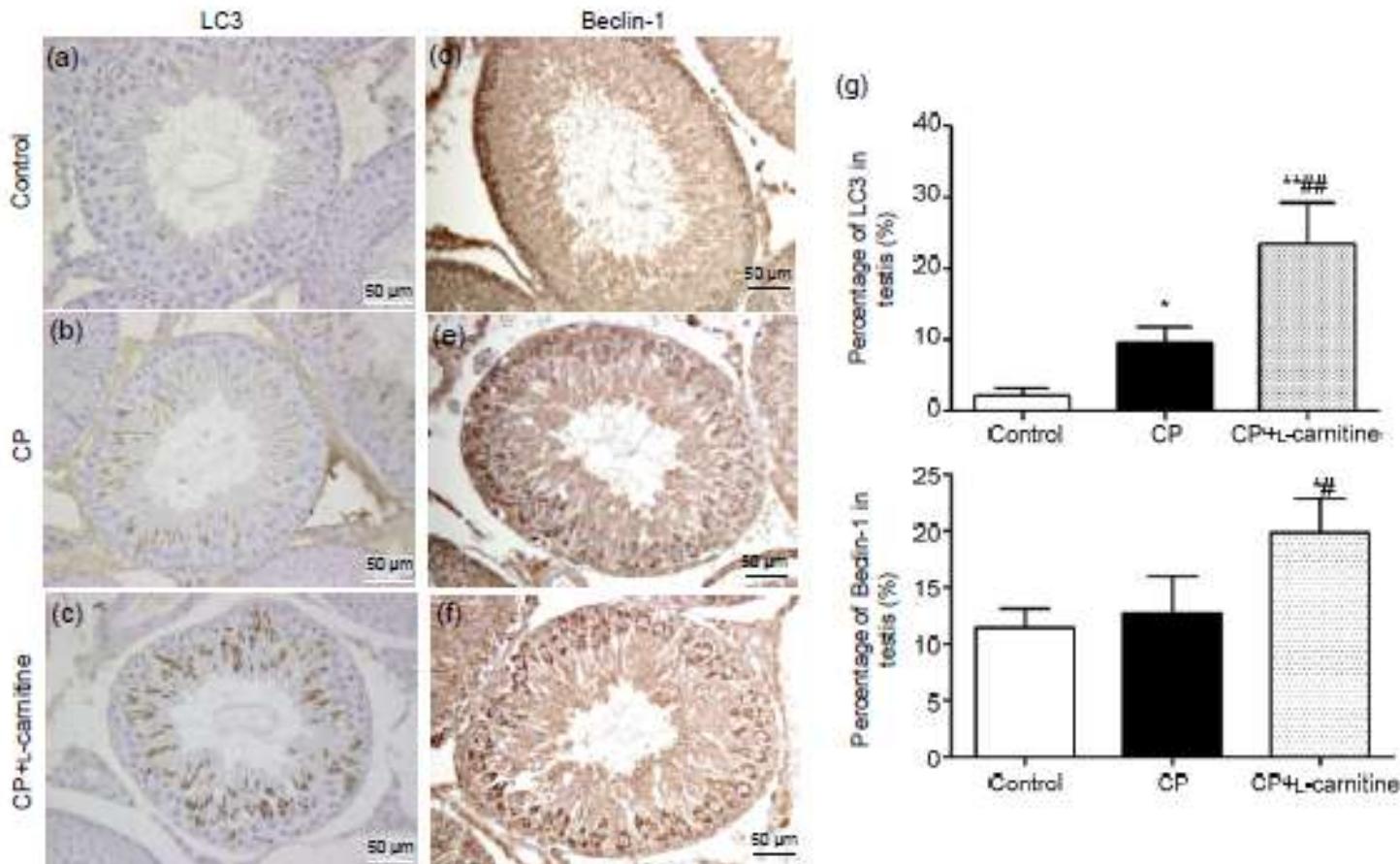


## TMBIM6 (transmembrane BAX inhibitor motif containing 6) Enhances Autophagy and Reduces Renal Dysfunction in a Cyclosporine A-induced nephrotoxicity model

Raj Kumar Yadav, Geum-Hwa Lee, Hwa-Young Lee, Bo Li, Han-Eul Jung, Harun-Or Rashid, Min Kyung Choi, Binod Kumar Yadav, Woo-Ho Kim, Kyung-Woon Kim, Byung-Hyun Park, Won Kim, Yong-Chul Lee, Hyung-Ryong Kim & Han-Jung Chae

Autophagy 2015, in press

# Autophagy: L-Carnitine



# Autophagy Regulation

# Autophagy: Regulation



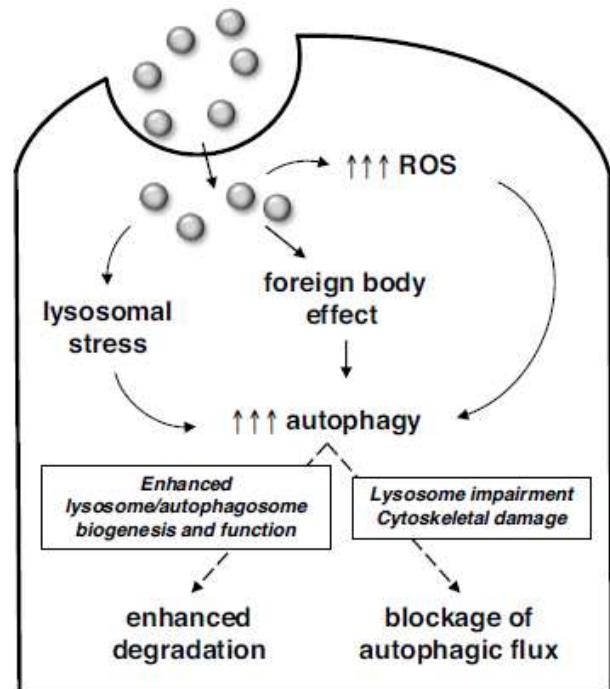
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Current Opinion in  
Biotechnology

**Differential autophagic responses to nano-sized materials**

Lauren Popp<sup>1</sup> and Laura Segatori<sup>1,2,3</sup>



Current Opinion in Biotechnology 2015, 36:129–136

# Autophagy: Therapeutic Modulation



## Autophagy and Transplant Immunity

Therapeutic compounds and targets that modulate autophagy

Compound	Effect on autophagy	Mechanism of action and target
Hydroxychloroquine	Inhibitor	Lysosomal acidification
3-MA, Wortmannin	Inhibitor	Class III PI3K
Anti-TNF $\alpha$	Inhibitor	Pro-autophagic cytokine block
P140 phosphopeptide	Inhibitor	Down-regulation of autophagic flux at the autolysosome stage
Tat-Beclin-1	Inducer	Interacts with a negative regulator of autophagy
Tensirolimus, sirolimus	Inducer	mTOR (incl. other actions)
Cyclosporine	Inducer	Mitochondrial permeability
Tamoxifen	Inducer	Beclin-1
Vitamin D	Inducer	mTOR inhibition
Bortezomib	Inducer	mTORC1 inhibition
Carbamazepine, valproate	Inducer	Inositol levels

3-MA, 3-methyladenine; PI3K, phosphatidylinositol 3-kinase; TNF, tumor necrosis factor; mTOR, mechanistic target of rapamycin; mTORC, mTOR complex.

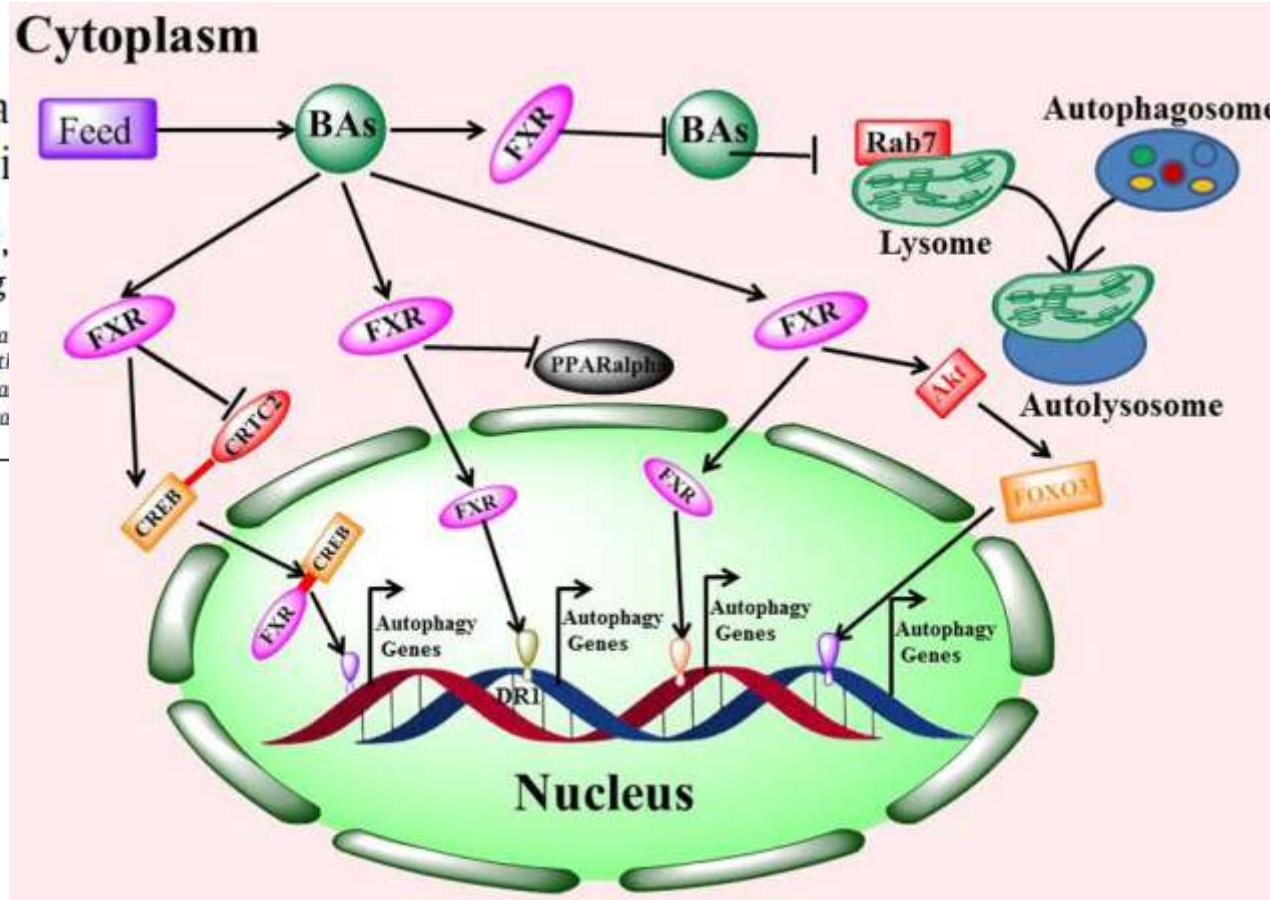
# Autophagy: Regulation

Review

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Zili Zhang<sup>a</sup>,  
Feng Zhang

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<sup>b</sup> Department of Pati  
<sup>c</sup> Department of Pha  
<sup>d</sup> Jiangsu Key Labora



# Autophagy: Regulation

Transcriptional regulation of autophagy.

Transcription factor	The effect of autophagy	The target genes	Conditions
FXR	Downregulation	<i>LCS, ATG4, ATG7, ATG10, Wipi1, Dfcp1, ULK1, LAMP2, P62, PI3KCIII, Bnip3</i>	Feeding or pharmacological activation
PPAR alpha	Upregulation	<i>ATG2, ATG4, ATG12, ATG16, Pink1, Bnip3, Wipi1, LC3, P13KCIII</i>	Fasting or pharmacological activation
PPAR beta/delta	Upregulation or downregulation	<i>ATG5, ATG7, LC3, Beclin1, P62, ULK1, Bnip3</i>	Depending on the different pathological conditions
PPAR gamma	Upregulation or downregulation	<i>ATG7, ATG12, LC3, P62, ULK1, LAM PI, BCL2, Beclin1, Pink1</i>	Depending on the different pathological conditions
TFEB	Upregulation	<i>ATG4, ATG9, BCL2, LC3, SQSTM1, Wipi1, UVARG</i>	Under different pathological conditions
NF-kappa B	Upregulation or downregulation	<i>BCL2, Bnip3, BECN1, SQSTM1</i>	Under different pathological conditions
HIF-1alpha	Upregulation	<i>Bnip3, BCL2, LC3, Beclin1, PI3KCIII</i>	Under different pathological conditions
P53	Upregulation or downregulation	<i>ATG2, ATG4, ATG7, ATG10, BCL2, ULK1, DRAM1, AMPK</i>	Basal levels: downregulation pathological conditions: upregulation
FOXO	Upregulation	<i>ATG8, ATG12, ATG4B, Gabarapl1, VSP34, BECLIN1</i>	Cytoplasm: downregulation Nucleus: upregulation
E2F	Upregulation or downregulation	<i>Bnip3, LC3, ULK1, DRAM, ATG1, ATG5</i>	Depending on the different pathological conditions
STAT	Downregulation	<i>ATG3, ATG12, BCL2, Bnip3, BECN1</i>	Cytoplasm: downregulation nucleus: downregulation
GATA	Upregulation or downregulation	<i>ATG4, ATG8, LC3, ATG12, Bnip3, ATG5, ATG7, BECN1</i>	CATA1: upregulation GATA4: downregulation

# Autophagy: Regulation



## The Pro-apoptotic STK38 Kinase Is a New Beclin1 Partner Positively Regulating Autophagy

Carine Joffre,<sup>1,5</sup> Nicolas Dupont,<sup>2</sup> Lily Hoa,<sup>3</sup> Valenti Gomez,<sup>3</sup> Raul Pardo,<sup>4</sup> Catarina Gonçalves-Pimentel,<sup>4</sup> Pauline Achard,<sup>5</sup> Audrey Bettoun,<sup>1</sup> Brigitte Meunier,<sup>1</sup> Chantal Bauvy,<sup>2</sup> Ilaria Cascone,<sup>1</sup> Patrice Codogno,<sup>2,\*</sup> Manolis Fanto,<sup>4,\*</sup> Alexander Hergovich,<sup>3,\*</sup> and Jacques Camonis<sup>1,\*</sup>

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<sup>2</sup>INSERM U1151-CNRS UMR 8253, Institut Necker Enfants-Malades, Paris 75993, France

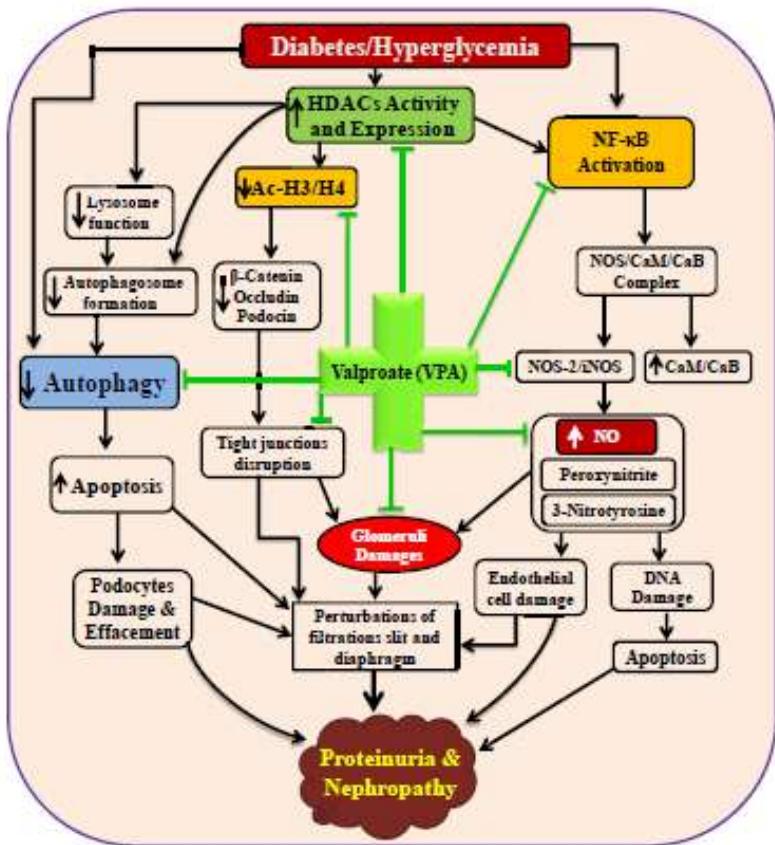
<sup>3</sup>University College London, Cancer Institute, London WC1E 6BT, UK

<sup>4</sup>Department of Basic and Clinical Neuroscience, Kings College London, London SE5 9NU, UK

<sup>5</sup>Cancer Research Center of Toulouse, UMR1037, Toulouse 31037, France

Current Biology 25, 1–14, October 5, 2015

# Autophagy: Regulation



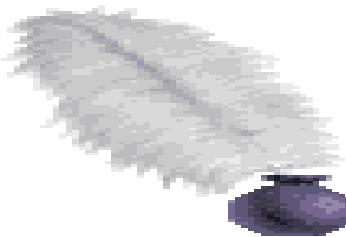


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**SUCCESS**  
belongs only to  
those who are  
willing to  
**work harder**  
than anyone  
else

